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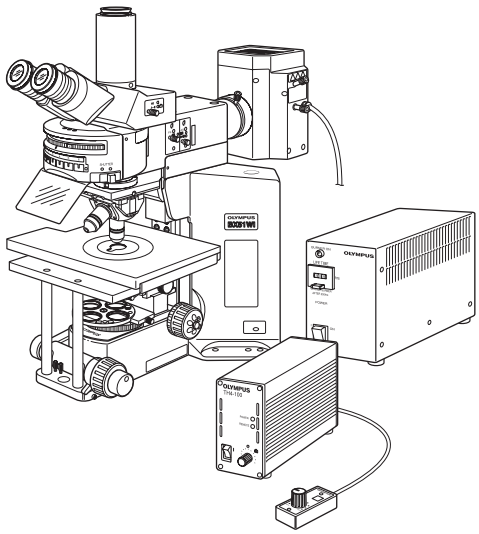

INSTRUCTIONS

BX51WI

FIXED-STAGE UPRIGHT MICROSCOPE

This instruction manual is for the Olympus Fixed-Stage Upright Microscope Model BX51WI. To ensure the safety, obtain optimum performance and to familiarize yourself fully with the use of this microscope, we recommend that you study this manual thoroughly before operating the microscope. Retain this instruction manual in an easily accessible place near the work desk for future reference.

This publication is printed on 100% recycled paper

AX7657

CONTENTS

Correct assembly and adjustments are critical for the microscope to exhibit its full performance. If you are going to assemble the microscope yourself, please read Chapter 9, "ASSEMBLY" (pages 36 to 42) carefully. For the modules provided with instruction manuals, also read the assembly procedures in their instruction manuals.

IMPORTANT – Be sure to read this section for safe use of the equipment. – 1-2

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■ **PROPER SELECTION OF THE POWER SUPPLY CORD** 43-44

This device complies with the requirements of directive 98/79/EC concerning in vitro diagnostic medical devices. CE marking means the conformity to the directive.

NOTE: This equipment has been tested and found to comply with the limits for a Class A digital device, pursuant to Part 15 of the FCC Rules. These limits are designed to provide reasonable protection against harmful interference when the equipment is operated in a commercial environment. This equipment generates, uses, and can radiate radio frequency energy and, if not installed and used in accordance with the instruction manual, may cause harmful interference to radio communications. Operation of this equipment in a residential area is likely to cause harmful interference in which case the user will be required to correct the interference at his own expense.

FCC WARNING: Changes or modifications not expressly approved by the party responsible for compliance could void the user's authority to operate the equipment.

IMPORTANT

This microscope employs a UIS2 (UIS) (Universal Infinity System) optical design, and should be used only with UIS2 (UIS) eyepieces, objectives and condensers for the BX2 series. (Some of the modules designed for the BX series are also usable. For details, please consult Olympus or the latest catalogues.)
To obtain comprehensive understanding on the operating procedures, please also read the separately provided instruction manuals.

Instruction manual	Contents
BX51WI	Explanation of transmitted brightfield observation, differential interference contrast observation and infrared observation
TH4	Explanation of the external halogen bulb power supply unit
BX-URA2/BX-RFA	Explanation of reflected light fluorescence observation
WI-DPMC	Explanation of the variable-magnification dual-port observation tube
WI-XYM/XYS	Explanation of the XY mover/bridge stage
WI-SSNP	Explanation of the swinging-sliding revolving nosepiece

⚠ SAFETY PRECAUTIONS

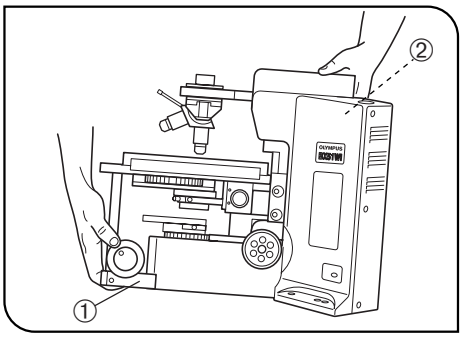


Fig. 1

1. After the equipment has been used in an observation of a specimen that is accompanied with a potential of infection, clean the parts coming in contact with the specimen to prevent infection.
   - Moving this product is accompanied with the risk of dropping the specimen. Be sure to remove the specimen before moving this product.
   - In case the specimen is damaged by erroneous operation, promptly take the infection prevention measures.
2. Culture liquid or water spilt on the stage, condenser or microscope may damage the equipment. Immediately wipe the liquid or water off if it is spilt on them.
3. When moving the microscope, disconnect the reflected light illuminator, observation tube and transmitted light lamp housing and carefully carry the microscope by the base (front edge) ① and the grasping part on the rear of the arm ② as shown in Fig. 1. (Weight: approx. 15 kg.)
   Also be careful against slipping of hands during carrying.

★ Damage to the microscope will occur if you grasp it by other parts including the stage, coarse/fine adjustment knob, etc.

Safety Symbols

The following symbols are found on the microscope. Study the meaning of the symbols and always use the equipment in the safest possible manner.

Symbol	Explanation
	Indicates that the surface becomes hot, and should not be touched with bare hands.
	Before use, carefully read the instruction manual. Improper use could result in personal injury to the user and/or damage to the equipment.

1 Getting Ready

1. A microscope is a precision instrument. Handle it with care and avoid subjecting it to sudden or severe impact.
2. The U-SWTR-2 super-widefield observation tube (FN 26.5) cannot be used with the BX51WI microscope.
3. The BX51WI microscope can be used with an intermediate attachment (such as a BX-URA2 or BX-RFA reflected light illuminator, U-ECA or U-CA magnification changer, etc.).
   Two intermediate attachments can be used only in the following conditions:
   - The U-CA or U-ECA magnification changer or U-FWO filter wheel can be mounted as the second attachment.
   - When a TV adapter with 1X or higher power is used, 2/3-inch CCD TV observation is possible.
   - The peripheral areas of the field of view may be obscured or cut off in binocular observation using the U-TR30-2, U-ETR or U-TR30IR (FN 22) super-widefield observation tube.
4. In IR (infrared) observation, the U-CA or U-ECA magnification changer can be used only when the U-ETR3 or U-TR30IR observation is used.
5. In photomicrography with visible light, correct exposure may be impossible if the microscope is set for IR observation. Be sure to engage the provided IR cut filter (light blue) before photomicrography.
6. When the XLUMPlanFI20XW objective is used, only the U-TV1X-2, U-TVCAC, U-PMTVC2XIR or U-PMTVC4XIR TV adapter can be used.
7. Do not attempt to remove or loosen the click springs and screws. Otherwise, Olympus can no longer warrant the performance of the microscope.
   The clicking force of the revolving nosepiece has been set weak in order to reduce vibrations during objective switching. To reproduce the correct click position, switch the objectives gently by operating the lever.
8. Caution for use of the U-ETR3 upright trinocular tube:
   When the aperture stop of the condenser is reduced using a reflected light fluorescence illuminator and the LUMPlanFI60XW objective, part of the observed field of view may be obscured slightly. This is due to the reduction of the light intensity in the field of view due to the narrow aperture and is not due to a defective optical adjustment of the microscope.
   This phenomenon does not affect the photomicrography or TV camera light path.

2 Maintenance and Storage

1. To clean the lenses and other glass components, simply blow dirty away using a commercially available blower and wipe gently using a piece of cleaning paper (or clean gauze).
   If a lens is stained with fingerprints or oil smudges, wipe it gauze slightly moistened with commercially available absolute alcohol.

⚠Since the absolute alcohol is highly flammable, it must be handled carefully.
Be sure to keep it away from open flames or potential sources of electrical sparks — for example, electrical equipment that is being switched on or off.
Also remember to always use it only in a well-ventilated room.

2. Do not attempt to use organic solvents to clean the microscope components other than the glass components. To clean them, use a lint-free, soft cloth slightly moistened with a diluted neutral detergent.
3. Never attempt to disassemble any part of the microscope.
4. When not using the microscope, make sure to set the main switch to "O" (OFF), confirm that the lamp housing is cool enough and cover the microscope with the provided dust cover.
5. When disposing of this unit, check the regulations and rules of your local government and be sure to observe them.

3 Warning Indication

A warning sticker is attached to a part where special precaution is required when handling and using the system. Always heed the warning.

Warning indication position	**Lamp housing (U-LH100-3/U-LH100IR) (Warning against high temperature)**

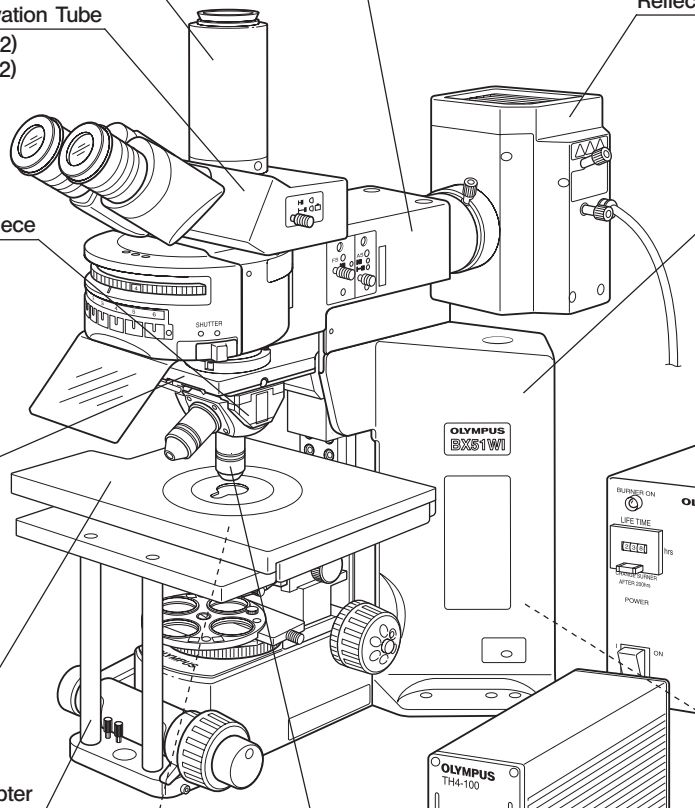
4 Caution

If the microscope is used in a manner not specified by this manual, the safety of the user may be imperiled. In addition, the equipment may also be damaged. Always use the equipment as outlined in this instruction manual.

The following symbols are used to set off text in this instruction manual.
▲: Indicates that failure to follow the instructions in the warning could result in bodily harm to the user and/or damage to equipment (including objects in the vicinity of the equipment).
★: Indicates that failure to follow the instructions could result in damage to equipment.
◎: Indicates commentary (for ease of operation and maintenance).

1 MODULE NOMENCLATURE

◎The modules shown below are only the representative modules. As there are other modules which can be combined with the microscope but are not shown below, please also refer to the latest Olympus catalogues or your dealer.



Transmitted Arm BX-ARM
or Reflected Light Fluorescence Illuminator
- BX-URA2
- BX-RFA

Straight Photo Tube U-SPT

Trinocular Observation Tube
- U-TR30-2 (FN 22)
- U-TR30IR (FN 22)
- U-ETR3 (FN 22)

Reflected Light Mercury Lamp Housing
- U-LH100HG
- U-LH100HGAPO

Microscope Frame BX51WIF

Revolving Nosepiece
- WI-SRE3
- WI-SNPXLU2
- U-SLRE

Power Supply Unit
(for 100 W Mercury Burner)

Revolving Arm
WI-NPA

Cross Stage
IX-SVL2
or commercially
available bridge
stage

Transmitted Light Lamp Housing
- U-LH100-3
- U-LH100IR

Fixed-Stage Adapter
WI-FSH

Power Supply Unit (for Halogen Bulb)
TH4

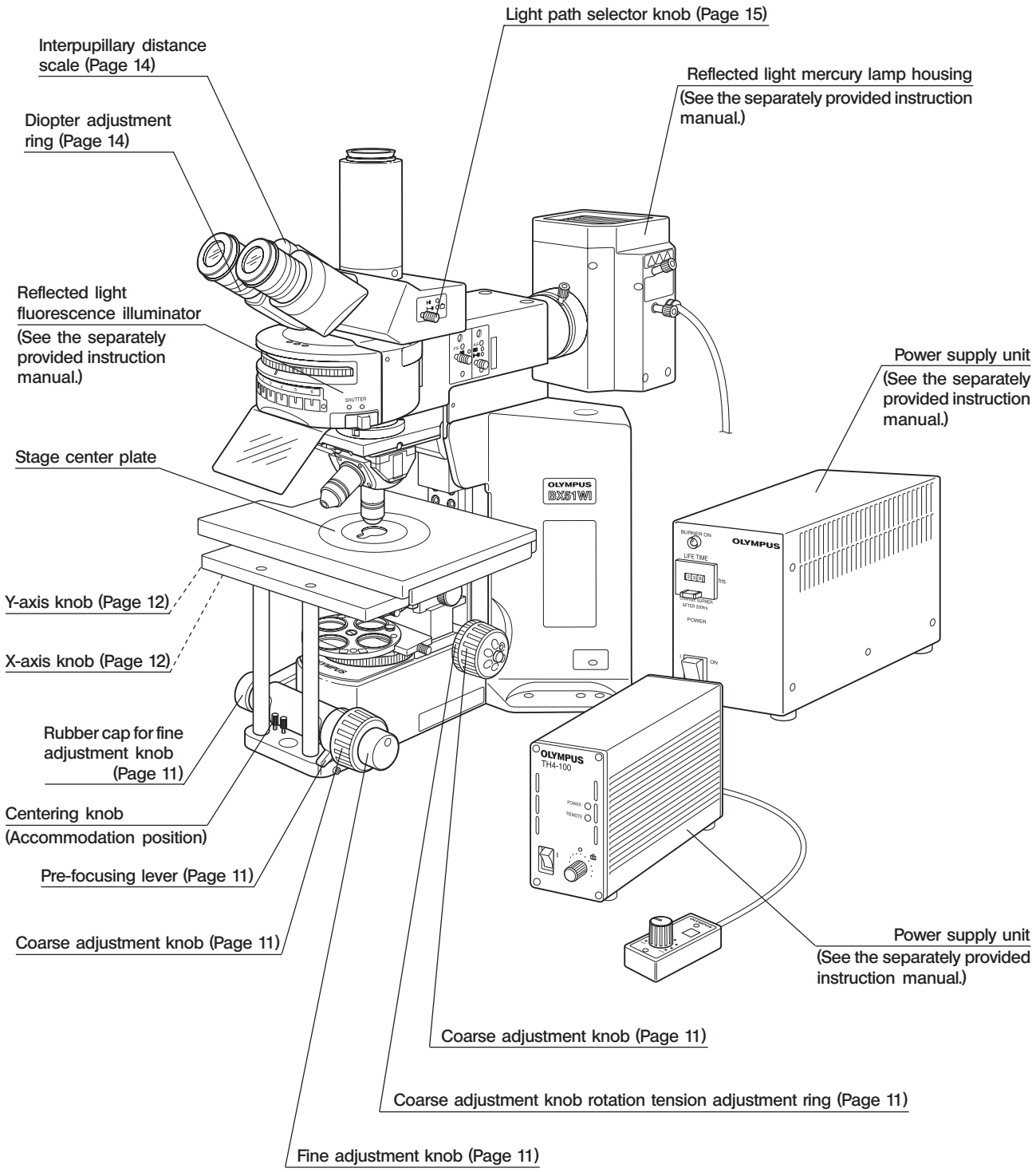
Hand Switch TH4-HS

Condenser
- WI-UCD
- WI-DICD
- WI-OBCD
- U-UCD8
- U-SC3
- U-AAC
- U-AC2

Objective
- MPLN5X
- MPlan5X
- UMPlanFI10XW/20XW
- LUMPlanFI40XW
- LUMPlanFI40XW/IR-2
- LUMPlanFI60XW
- LUMPlanFI60XW/IR-2
- LUMPlanFI100XW
- XLUMPlanFI20XW
  (exclusively for use with the WI-SNPXLU2)
- XLFluor2X/340
  (exclusively for use with the U-SLRE)
- XLFluor4X/340
  (exclusively for use with the U-SLRE)

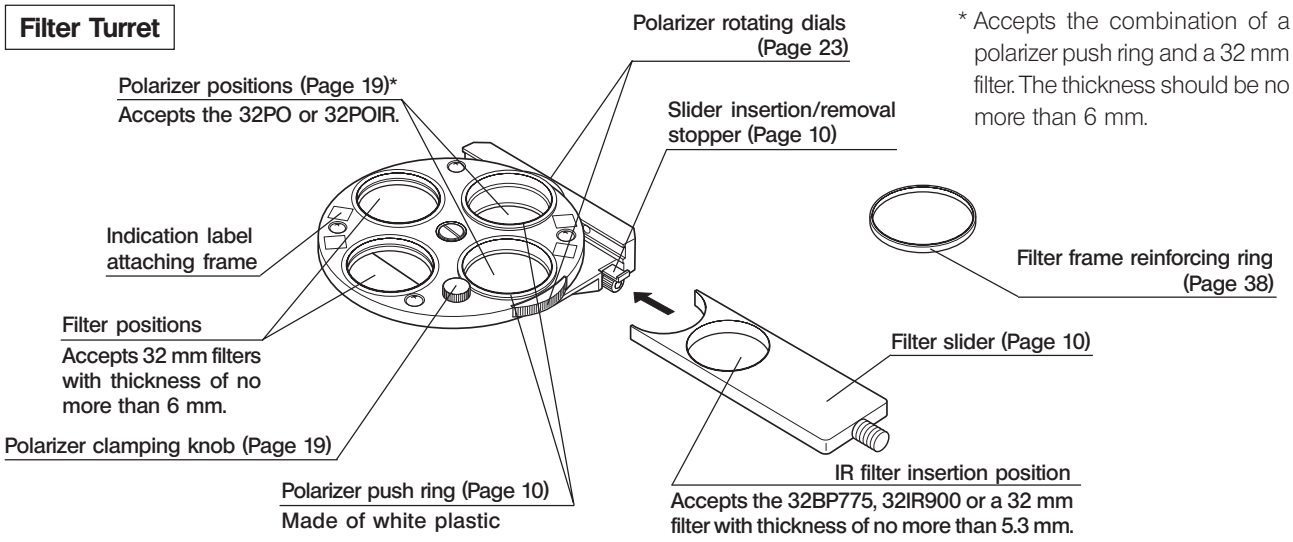
2 CONTROLS

◎If you have not yet assembled the microscope, read Chapter 9, "ASSEMBLY" (pages 36 to 42).


Light path selector knob (Page 15)
Interpupillary distance scale (Page 14)
Reflected light mercury lamp housing (See the separately provided instruction manual.)
Diopter adjustment ring (Page 14)
Reflected light fluorescence illuminator (See the separately provided instruction manual.)
Power supply unit (See the separately provided instruction manual.)
Stage center plate
OLYMPUS BX51WI
OLYMPUS
Y-axis knob (Page 12)
X-axis knob (Page 12)
Rubber cap for fine adjustment knob (Page 11)
OLYMPUS TH4-100
Centering knob (Accommodation position)
Pre-focusing lever (Page 11)
Coarse adjustment knob (Page 11)
Power supply unit (See the separately provided instruction manual.)
Coarse adjustment knob (Page 11)
Coarse adjustment knob rotation tension adjustment ring (Page 11)
Fine adjustment knob (Page 11)

◎The descriptions on the filter turret, revolving nosepiece, condenser, etc. will be given in the subsequent pages.

Filter Turret

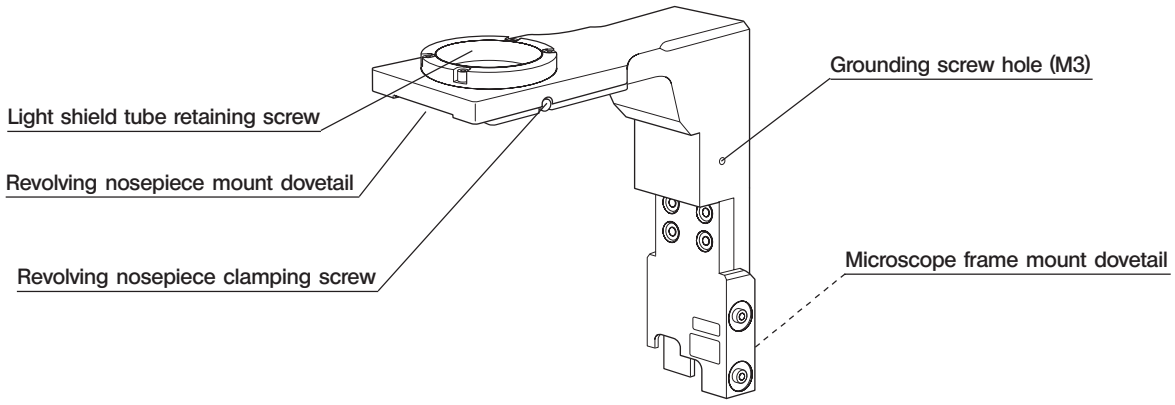

Polarizer rotating dials (Page 23)
Polarizer positions (Page 19)*
Accepts the 32PO or 32POIR.
Slider insertion/removal stopper (Page 10)
Indication label attaching frame
Filter frame reinforcing ring (Page 38)
Filter positions
Accepts 32 mm filters with thickness of no more than 6 mm.
Filter slider (Page 10)
Polarizer clamping knob (Page 19)
IR filter insertion position
Accepts the 32BP775, 32IR900 or a 32 mm filter with thickness of no more than 5.3 mm.
Polarizer push ring (Page 10)
Made of white plastic

* Accepts the combination of a polarizer push ring and a 32 mm filter. The thickness should be no more than 6 mm.

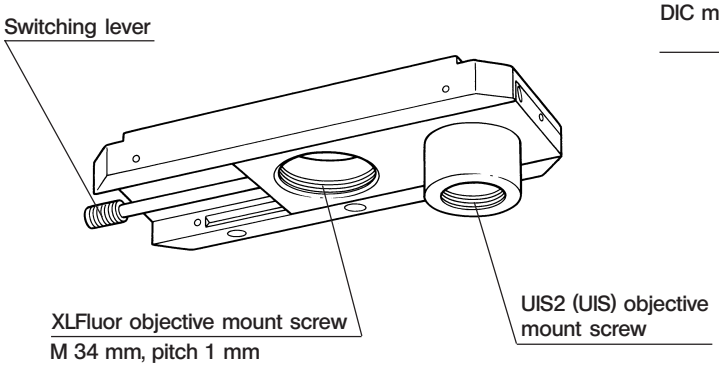
Revolving Arm WI-NPA

★Note that the revolving arm can be mounted only before the reflected light illuminator is mounted or before the transmitted light arm and IX-SVL2 stage are mounted (page 38).

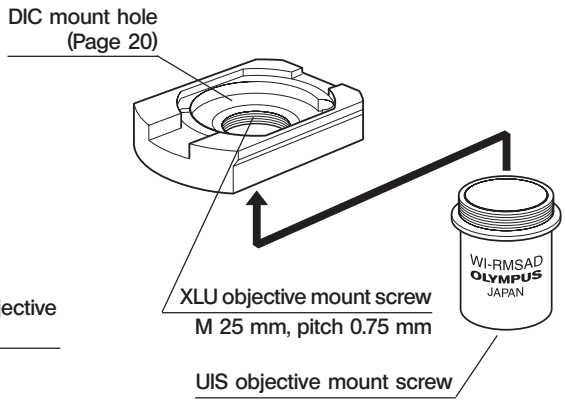
◎This revolving arm accepts the U-SLRE, WI-SNPXLU2 or WI-SRE3 revolving nosepiece.


Grounding screw hole (M3)
Light shield tube retaining screw
Revolving nosepiece mount dovetail
Microscope frame mount dovetail
Revolving nosepiece clamping screw

Sliding Revolving Nosepiece U-SLRE

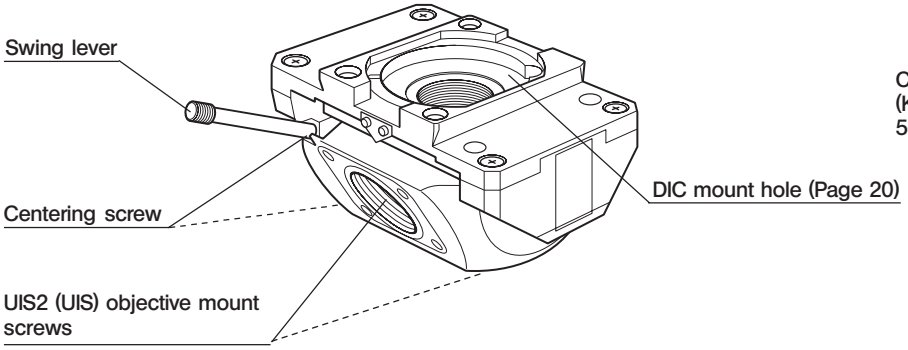


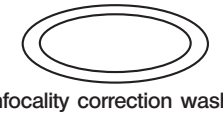
Single-Position Revolving Nosepiece XLU WI-SNPXLU2



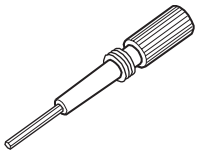
RMS Adapter WI-RMSAD

Swinging Revolving Nosepiece WI-SRE3



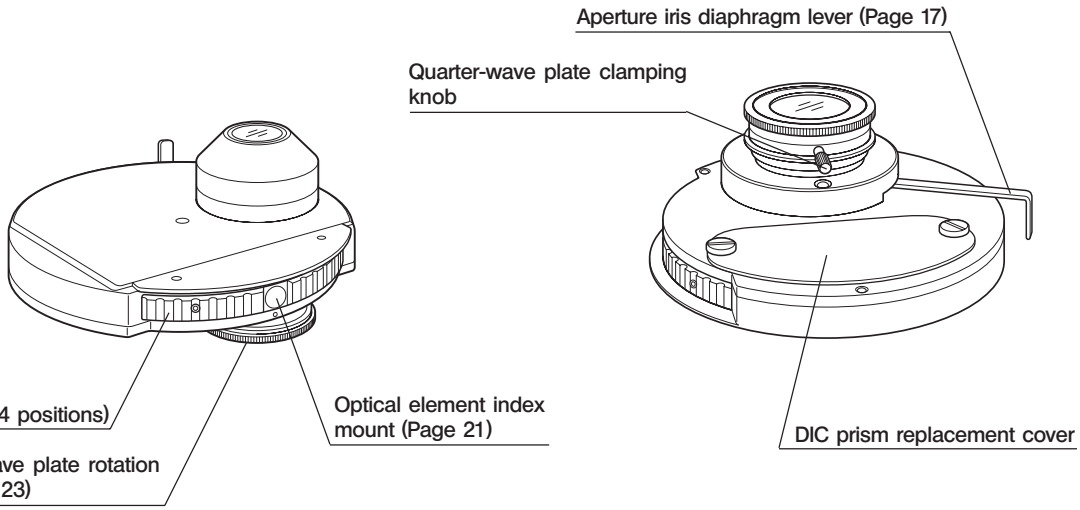


Confocality correction washers (Kinds of thickness: 10, 30 and 50 µm; three washers per kind)

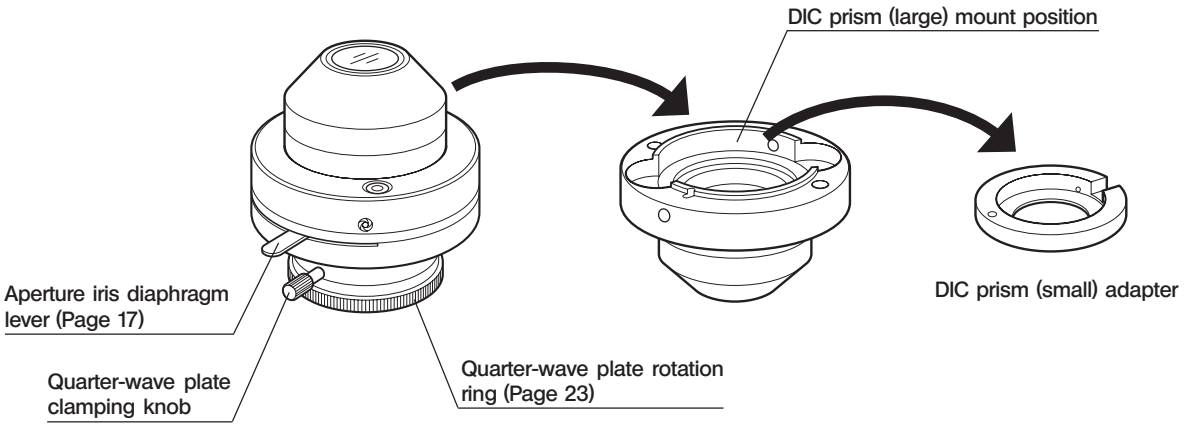


Centering knob

Long-WD Universal Condenser WI-UCD

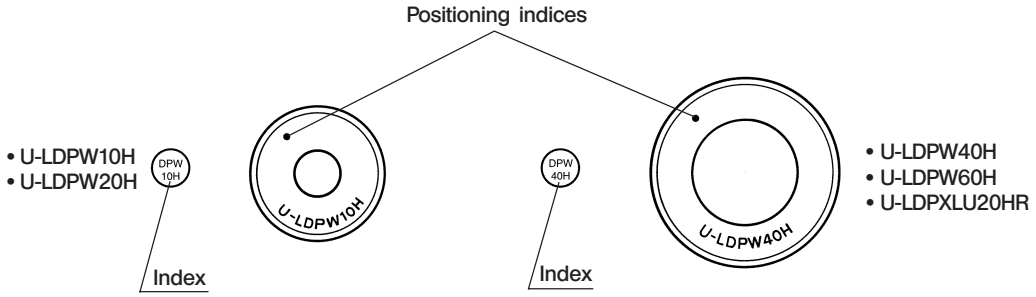

Aperture iris diaphragm lever (Page 17)
Quarter-wave plate clamping knob
Turret (4 positions)
Optical element index mount (Page 21)
Quarter-wave plate rotation ring (Page 23)
DIC prism replacement cover

Long-WD DIC Condenser WI-DICD

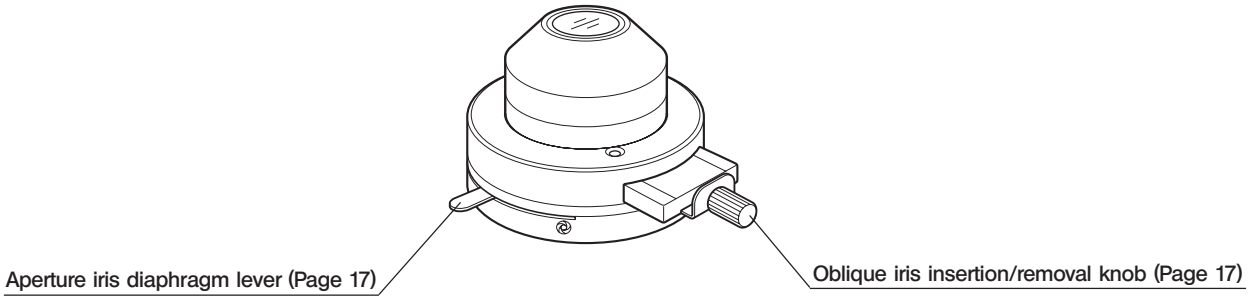

DIC prism (large) mount position
Aperture iris diaphragm lever (Page 17)
Quarter-wave plate clamping knob
Quarter-wave plate rotation ring (Page 23)
DIC prism (small) adapter

Differential Interference Contrast Prisms (For Condenser)

◎The WI-UCD condenser accepts two large and two small DIC prisms while the WI-DICD condenser accepts one large or small DIC prism.
When selecting the brightfield (BF) light path using the WI-UCD, leave one DIC prism (large) mount position empty.



Long-WD Oblique Condenser WI-OBCD

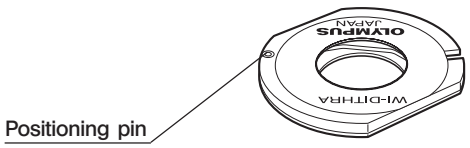


High-Resolution DIC Prism A WI-DICTHRA2
DIC Prism WI-DICT2

◎This prism can be mounted in the DIC prism position of the WI-SNPXLU2 or WI-SRE3.

Applicable condensers

WI-DICTHRA2: WI-UCD, WI-DICD
WI-DICT2: U-UCD8



■ Condensers and Applicable Objective Magnifications

Condenser	Applicable Objective Magnification
WI-UCD WI-DICD WI-OBCD	5X or more
U-UCD8 U-SC3	2X or more
U-AAC	10X or more
U-AC2	5X or more

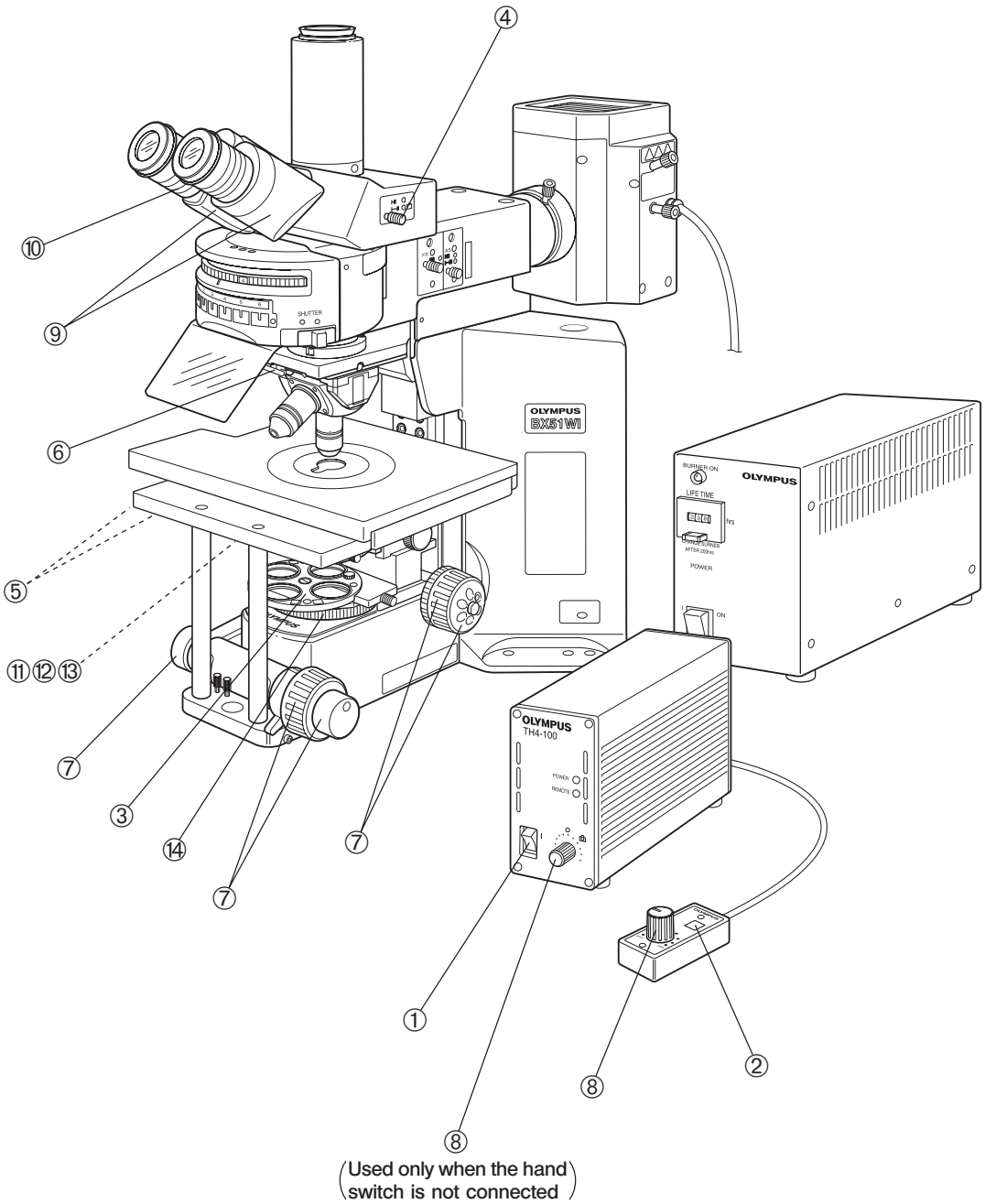
3 TRANSMITTED LIGHT BRIGHTFIELD OBSERVATION PROCEDURE

◎The following flow shows the operating procedure for the transmitted light brightfield observation which is the basic observation method of this microscope. The operating procedures for DIC observation, fluorescence DIC observation and IR DIC observation will be described separately in Chapter 5, "OTHER OBSERVATION METHODS" on page 19.

	(Controls Used)	(Page)
Set the main switch to " I " (ON).	①Main switch	(P. 10)
	②Lamp ON-OFF switch (Only when the hand switch is used)	(P. 10)
Disengage the filter from the light path.	③Filter turret	(P. 10)
Select the binocular light path.	④Light path selector knob	(P. 15)
Place the specimen on the stage.	⑤X-axis and Y-axis knobs	(P. 12)
Engage a low-magnification objective in the light path.	⑥Swing lever	(P. 13)
Bring the specimen in focus.	⑦Coarse/fine adjustment knobs	(P. 11)
Adjust the brightness.	⑧Light intensity control knob	(P. 10)
Adjust the interpupillary distance.	⑨Binocular tube	(P. 14)
Adjust the diopter.	⑩Diopter adjustment ring	(P. 14)
Adjust the light axis.	⑪Condenser height adjustment knob	(P. 16)
	⑫Condenser centering knob	(P. 16)
Adjust the aperture iris and field iris diaphragms.	⑬Aperture iris diaphragm lever	(P. 17)
	⑭Field iris diaphragm ring	(P. 16)
Engage the objective to be used in the light path and bring the specimen in focus.	⑥Swing lever	(P. 13)
	⑦Coarse/fine adjustment knobs	(P. 11)
Engage the required filters.	③Filter turret	(P. 10)
Adjust the brightness.	⑧Light intensity control knob	(P. 10)
Start observation.		

Tip for microscope operation

In patch-clamp testing, switch the microscope controls cautiously and gently so that the patch electrodes do not slip off.



© Make a photocopy of the observation procedure pages and post it near your microscope.

4 USING THE CONTROLS

4-1 Microscope Base, Power Supply Unit (TH4)

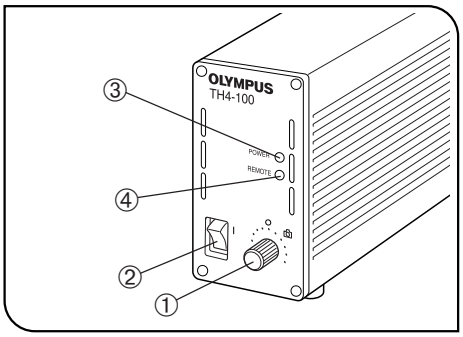


Fig. 2

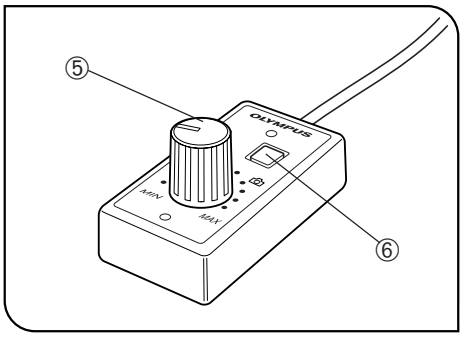


Fig. 3

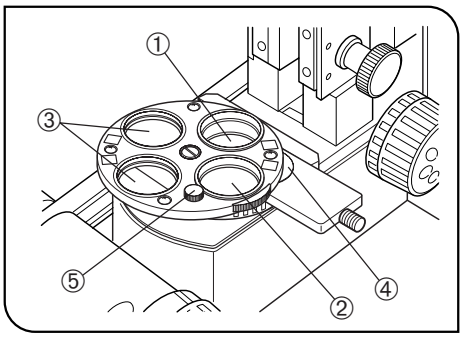


Fig. 4

1 Controlling the Light Intensity (TH4) (Figs. 2 & 3)

◎See the separately provided instruction manual for details.

1. Make sure that the light intensity control knob ① is set to MIN (minimum voltage), then set the main switch ② to " I " (ON). (The POWER LED ③ should light up.)
2. Turn the knob ① clockwise toward MAX (maximum voltage) to increase the intensity and brightness.

◎The 📷 marking indicates the position where the optimum daylight for color photography is obtained when the LBD filter is engaged in the light path.

Operation Using the Hand Switch

◎When the hand switch is connected (when the REMOTE LED ④ is lit), the light intensity control knob ① is defeated and only the light intensity control knob ⑤ of the hand switch can be used.
The hand switch is provided with double-side adhesive tape so that it can be attached onto a convenient position for operation.

1. After setting the main switch ② to " I " (ON), press the lamp ON-OFF switch ⑥ to ON and adjust the brightness with the intensity control knob ⑤.
2. To turn the lamp OFF, press the lap ON/OFF switch ⑥ again to OFF.

★The lighting of the REMOTE LED ④ indicates that the hand switch is standing by. The hand switch consumes a power of about 2.5 W when it stands by.
When the system is not to be used for a lone period, be sure to set the main switch ② to "O" (OFF).

2 Using the Filter Turret (Fig. 4)

◎Filters with a diameter of 32 mm can be inserted in positions ① to ④.

1. Filter positions ① and ② are rotatable. When the 32PO polarizer or 32POIR polarizer is placed in either position, the polarizer or filter can be fixed by using the push ring (made of white plastic).

◎When filter position ① is engaged in the light path, the rotation fixing knob ⑤ comes at the front where the operation is easy.

2. Filter position ③ accepts any type of 32 mm filter.

★When using two filters together, the thickness of the lower filter should be no more than 2 mm. Otherwise, the upper filter may drop during rotation.

3. Filter position ④ accepts the 32BP775 or 32IR900 filter. As the filter cannot be inserted unless the filter slider is removed, remove it by releasing the insertion/removal stopper below the slider and loosening the slider clamping screw using the provided Allen screwdriver.

4-2 Focusing Block

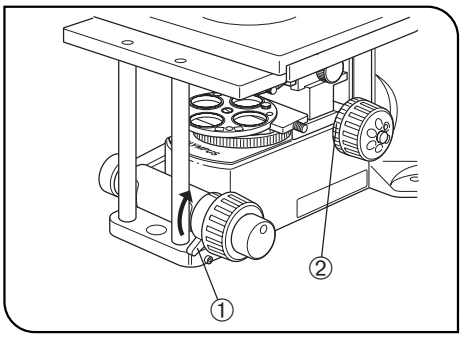


Fig. 5

1 Using the Pre-focusing Lever (Fig. 5)

◎The pre-focusing lever prevents collision between the specimen and objective and simplifies the focusing operation. After bringing the specimen into approximate focus with the coarse adjustment knob, turn the pre-focusing lever ① in the direction of the arrow to lock it. Hereafter, the lower limit of the coarse adjustment will be limited at the position where the lever is locked. When bringing a specimen in focus, approximate focus can be obtained by simply lowering the coarse adjustment to the stopper position so all you have to do more is control the fine adjustment knob.

◎The up/down movement using the fine adjustment knob is not limited.

★When the pre-focusing lever is locked, the coarse adjustment stroke is limited by the mechanism and it cannot reach the previous upper limit. If you want to control the coarse adjustment knob to the previous upper limit, unlock the pre-focusing lever.

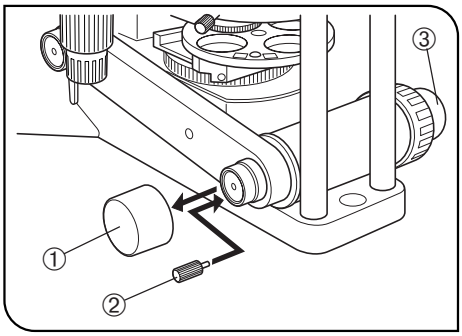


Fig. 6

2 Using the Fine Adjustment Fast-Feed Knob (Fig. 6)

◎Fine focusing is usually possible while the rubber cap ① is attached. However, when it is desirable to allow the fine adjustment knob to vary focusing by a large amount, though this is not be as large as with the coarse adjustment knob, the rubber cap can be removed and the provided fine adjustment fast-feed knob attached in place.

◎If you remove the knob by loosening the screw clamping the fine adjustment knob ③ from the opposite side, the fine adjustment can be controlled using the tip or thick of your finger.

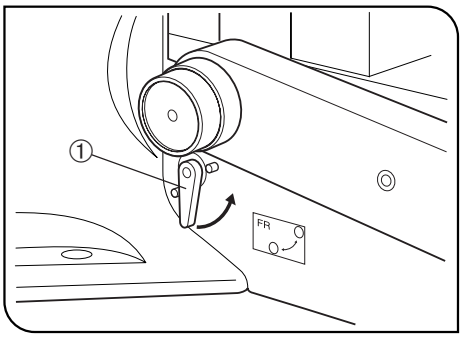


Fig. 7

3 Using the Frost Switching Lever (Fig. 7)

◎Low observation light can be brightened by turning the frost switching lever ① which controls the built-in frost filter, in the direction of the arrow. However, although the brightness is increased, irregularity in lighting may also increase.

4 Adjusting the Coarse Adjustment Knob Rotation Tension (Fig. 5)

★Do not adjust the coarse adjustment knob rotation tension adjustment ring (② in Fig. 5) because the belt interlocking of the ring with the coarse adjustment knob on the front has been adjusted at the factory. If the tension is varied, the accuracy of the pre-focusing lever will deteriorate.

4-3 Stage (IX-SVL2)

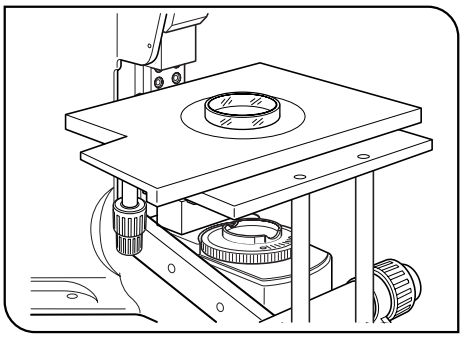


Fig. 8

1 Placing the Specimen (Fig. 8)

1. Place the specimen on the center of the stage.

◎The optional stage center plate (IX-CP50) makes it possible to observe a wide range of a big petri dish, etc. (Central hole diameter: 50 mm)

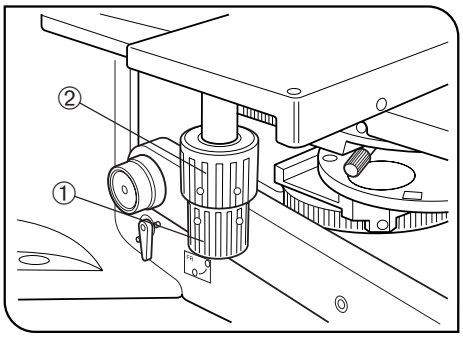


Fig. 9

2 Moving the Specimen (Fig. 9)

1. The specimen can be moved by turning the X-axis knob ① and Y-axis knob ②.
   The movement strokes are 50 mm (X-axis) x 43 mm (Y-axis).

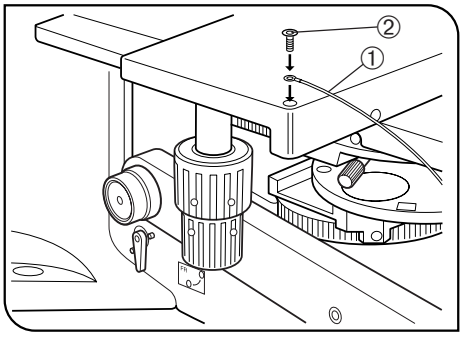


Fig. 10

3 Setting the Grounding (Fig. 10)

◎In case of electrical physiological experiment, etc., the specimen can be grounded from the stage.
Prepare a grounding wire ① and M4 screw ② and attach grounding as shown in Fig. 10.

★**The screw hole may sometimes be stuck by paint, etc. In such a case, screw in the M4 screw a few times to expose the metallic thread inside the screw hole and improve the contact before attaching the grounding wire firmly.**

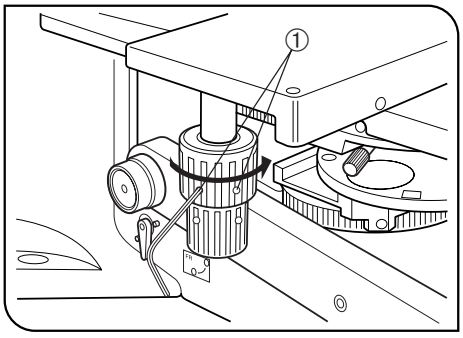


Fig. 11

4 Adjusting the X-Axis/Y-Axis Knob Rotation Tension (Fig. 11)

◎The rotation tension of the X-axis and Y-axis knobs can be adjusted independently.

1. Loosen the 2 set screws ① of a knob using the provided Allen wrench, hold the stage so that it will not move, then turn the knob to adjust the tension. Turning it in the direction of the arrow increases the tension and turning in the opposite direction decreases the tension.
2. After adjustment, tighten the set screws firmly.

★**If the tension of a knob is too heavy or too light, skipping or returning of image may occur during the stage movement.**

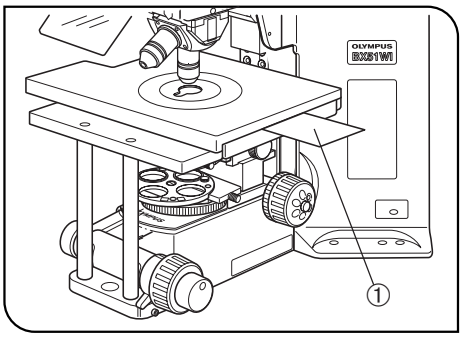


Fig. 12

5 Using the Light Shield Sheet (Fig. 12)

★**The light shield sheet provided with the reflected light fluorescence illuminator is too small to be used with the BX51WI. Always use the light shield sheet provided with the BX51WI microscope frame.**

◎During fluorescence observation using a low-magnification objective, the fluorescence image may be deteriorated due to light reflected from the condenser or the surroundings. In this case, use the light shield sheet.

1. Lower the condenser to the lower limit position using the condenser height adjustment knob.
2. Insert the light shied sheet all the way into the gap between the upper and lower stages on the side of the stage (IX-SVL2).

★**If the condenser is lowered insufficiently, the sheet cannot be inserted into the normal position and the light shielding effect cannot be obtained.**

6 Lowering the Stage Height

◎The stage can be lowered by 50 mm by removing the condenser holder.
See page 42 for details.

4-4 Revolving Nosepiece

⚠**If the petri dish in use is filled with liquid, it may splash when the objective is switched. As such liquids are sometimes toxic, be sure to move the revolving nosepiece away from the petri dish before switching the objective.**
Even after the revolving nosepiece has been moved, re-focusing is easy by making use of the pre-focusing lever (page 11).

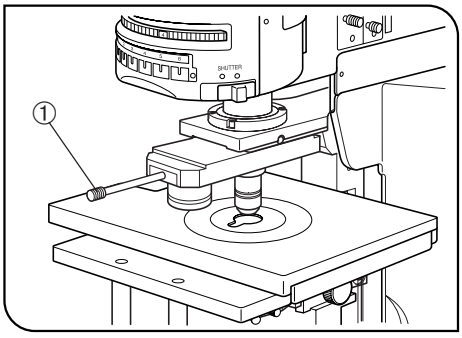


Fig. 13

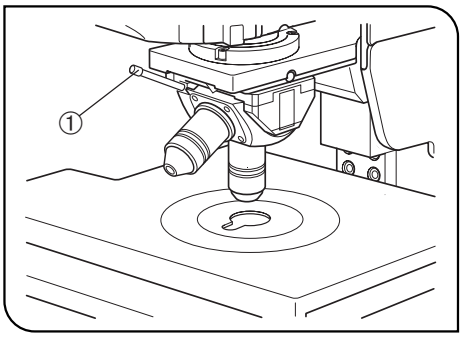


Fig. 14

1 Switching the Objectives (U-SLRE, WI-SRE3) (Figs. 13 & 14)

◎The clicking force of the revolving nosepiece has been set weak in order to reduce vibrations during objective switching.
To reproduce the correct click position, switch the objectives gently by operating the lever.

Sliding Revolving Nosepiece U-SLRE

Switch the objective by holding the objective switching lever ① and gently moving it back and forth.

◎By attaching the objective switching lever ① on the opposite side, a UIS objective can be positioned on the front side of the microscope.

Sliding Revolving Nosepiece WI-SRE3

Switch objectives by gently puling up or pushing down the swing lever ①.
Pull up or push down the swing lever gently until it hits the revolving nosepiece's stopper.

4-5 Observation Tube

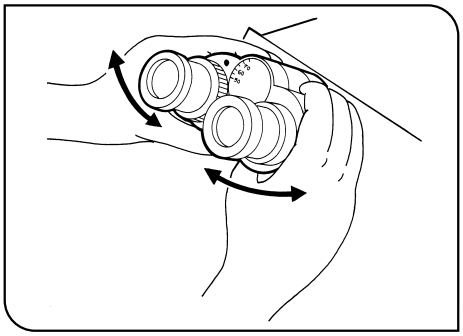


Fig. 15

1 Adjusting the Interpupillary Distance (Fig. 15)

While looking through the eyepieces, adjust for binocular vision until the left and right fields of view coincide completely. The index dot • indicates the interpupillary distance.

◎Note your interpupillary distance so that it can be quickly duplicated.

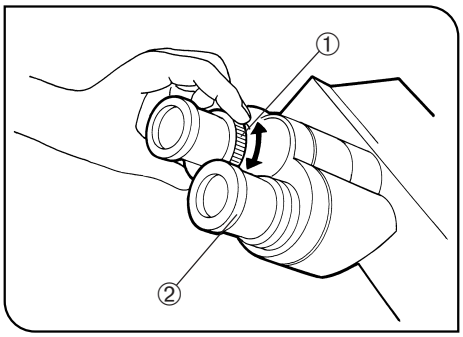


Fig. 16

2 Adjusting the Diopter (Figs. 16 & 17)

1. Looking through the eyepiece without the diopter adjustment ring, rotate the coarse and fine adjustment knobs to bring the specimen into focus.
2. Looking through the eyepiece sleeve with the diopter adjustment ring ①, turn only the ring to focus on the specimen. (Fig. 16)

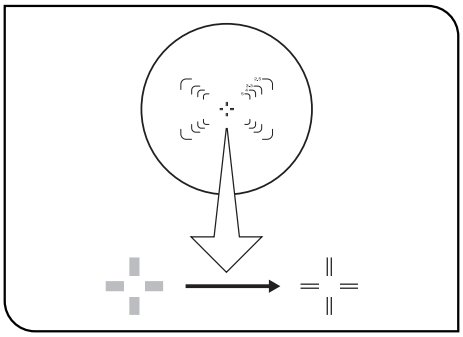


Fig. 17

Using a Finder Eyepiece

1. Looking through the right eyepiece with your right eye, turn the top of the eyepiece ② until a clearly defined double crosslines can be seen in the field of view. (Figs. 16 & 17)
2. Looking through the right eyepiece, rotate the coarse and fine adjustment knobs to bring the specimen and double crosslines into simultaneous focus.
3. Looking through the left eyepiece with your left eye, turn the diopter adjustment ring ① to focus on the specimen.

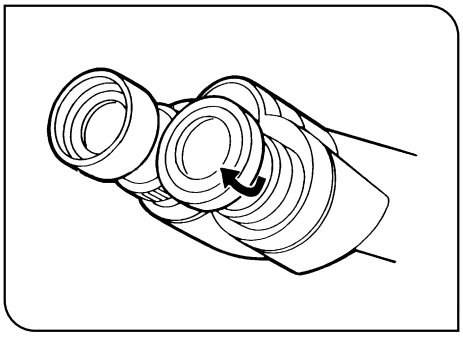


Fig. 18

3 Using the Eye Shades (Fig. 18)

When Wearing Eyeglasses

Use with the eye shades in the normal, folded-down position. This will prevent the eyeglasses from being scratched.

When Not Wearing Eyeglasses

Extend the folded eye shades in the direction of the arrow to prevent extraneous light from entering between the eyepieces and eyes.

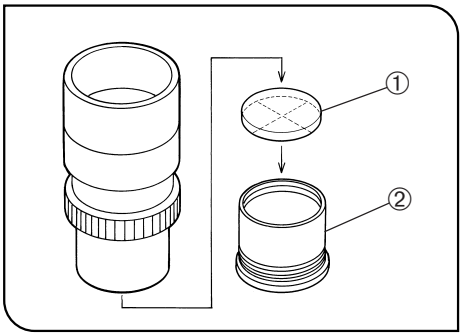


Fig. 19

4 Using Eyepiece Micrometer Disks (Fig. 19)

Eyepiece micrometer disks can be inserted into the WHN10X-H (or WHN10X) eyepieces.
However, if the eyepiece does not have the helicoid adjustment facility and your eyesight is poor, you may have difficulties in focusing on the eyepiece micrometer disk. In this case, it is recommended to look into the eyepiece through your eyeglasses.
Use 24 mm dia. x 1.5 mm micrometer disks.
Following Fig. 19, remove the micrometer mounting frame ② from the eyepiece and place a micrometer disk ① into the mounting frame.
Re-attach the micrometer mounting frame in the original position.

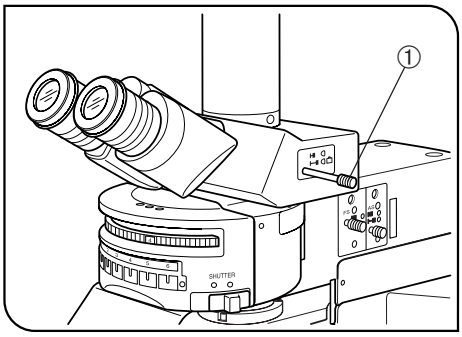


Fig. 20

5 Selecting the Light Path of Trinocular Tube (Fig. 20)

Slide the light path selector knob ① to select the desired light path.

Trinocular Tube	Light Path Selector Position		
	Pushed In	Intermediate	Pulled Out
U-TR30-2	Binocular 100%	Binocular 20%, TV, photo 80%	TV, photo 100%
U-ETR3	Binocular 100%		TV, photo 100%
U-TR30IR	Binocular 100%	Shutter	TV, photo 100%

4-6 Condenser

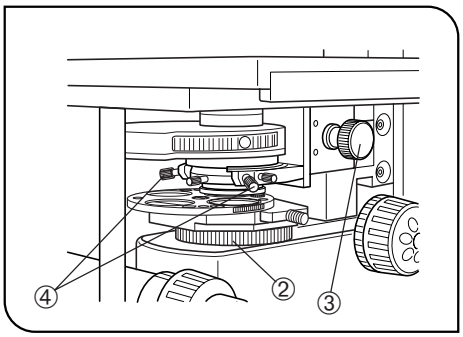


Fig. 21

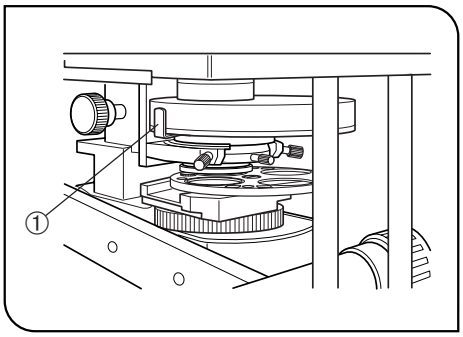


Fig. 22

1 Centering the Condenser (Figs. 21 & 22)

1. Set the aperture iris diaphragm lever ① to the open position. (Fig. 22)
2. Set the field iris diaphragm ring ② to the open position (→ ○). (Fig. 21)
3. Focus on the specimen using the 10X objective.
4. Close the field iris diaphragm ring ② so that the diaphragm image comes inside the field of view.
5. Manipulate the condenser height adjustment knob ③ to focus on the diaphragm image.
6. While opening the field iris diaphragm gradually, turn the two condenser centering screws ④ on the condenser holder to move the iris diaphragm image to the center of the field of view. (Fig. 21, Fig. A → Fig. B).
7. Gradually open the field iris diaphragm. The condenser is properly centered if the iris image is centered and inscribed in the field of view (Fig. B → Fig. C).

◎During actual use, open the field diaphragm slightly until its image circumscribes the field of view.

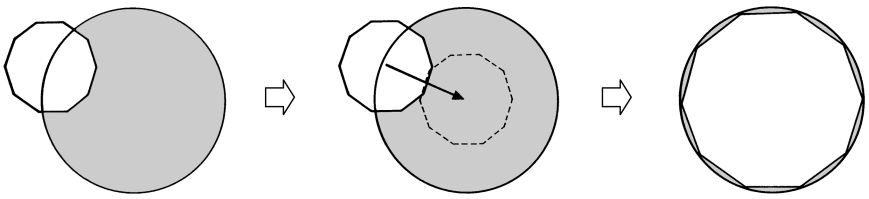


Fig. A Fig. B Fig. C

Field Iris Diaphragm

The field iris diaphragm restricts the diameter of the beam of light entering the objective and thus excludes extraneous light, improving image contrast. The diameter of the field iris should be adjusted for objective magnification to the extent that it just circumscribes the field of view.

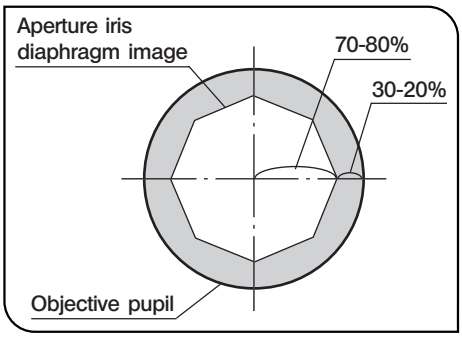


Fig. 23

Aperture Iris Diaphragm

- The aperture iris diaphragm determines the numerical aperture of the illumination system. Matching the numerical aperture of the illumination system with that of the objective provides better image resolution and contrast, and also increases the depth of focus.
- Since the contrast of microscope specimens is ordinarily low, setting the condenser aperture iris diaphragm to between 70% and 80% of the NA of the objective in use is usually recommended. If necessary, adjust the ratio by removing the eyepieces and looking into the eyepiece sleeve while adjusting the aperture iris diaphragm lever ① until the image shown in Fig. 23 is seen. (Fig. 22)

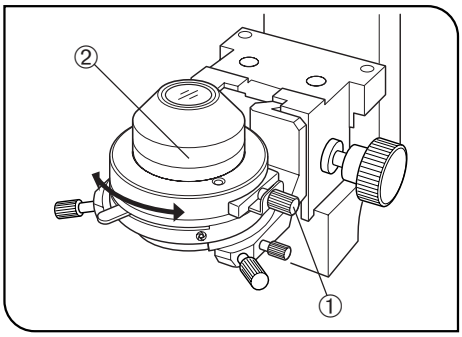


Fig. 24

2 Oblique Illumination (WI-OBCD) (Figs. 24 & 25)

◎The shading and 3D feeling of the specimen can be adjusted by varying the width and orientation of the area subjected to oblique illumination. This is possible with objectives from 5X to 100X.

★When the XLUMPlanFI20XW objective is used, the effects of oblique illumination cannot be manifested fully due to the high NA (0.95) of the objective.

Important

The oblique illumination presupposes that the field iris image is focused correctly.
Before proceeding to the following, pull out the oblique iris insertion/removal knob ① and bring the field iris image in focus (this is the same operation as that described on page 16).

1. Push in the oblique iris insertion/removal knob ①.
2. Turn the knob ① to adjust the width of the area illuminated by oblique illumination. (Fig. 25)
3. Adjust the orientation of the oblique illumination by turning the top part ② of the condenser.

◎Pull out the oblique iris insertion/removal knob when using the condenser as usual.
The width of the oblique illumination area is maintained even after the insertion/removal knob ① has been pulled out, so the same condition can be reproduced the next time the knob is pushed in.

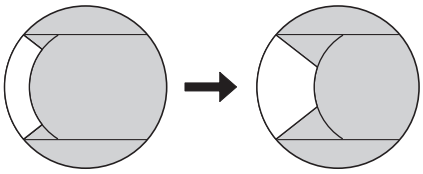


Fig. 25

4-7 Immersion Objectives

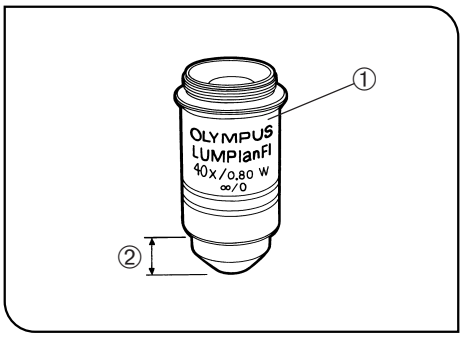


Fig. 26

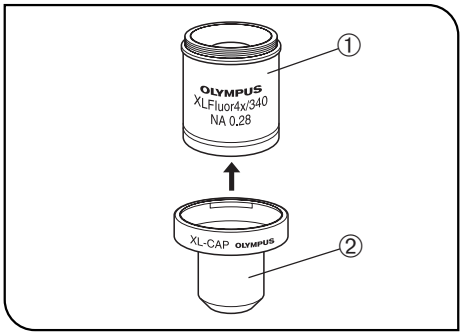


Fig. 27

1 Using Water Immersion Objectives (Figs. 26 & 27)

◎When the UMPlanFl-W series, LUMPlanFl-W series or XLUMPlanFl20XW objective is used, cultured tissue specimens which are often very thick can be observed by immersing the specimen, objective front lens and manipulator extremity in a medium with the same refractive index (water).

★**The electrically insulated area and immersion depth of the objective ① are shown by the range of ②.**

CAUTION **Do not immerse the entire objective, for this will cause malfunction.**
After every immersed use, be sure to clean the front lens with neutral detergent.

Water Immersion Cap for XL Objectives XL-CAP

In photometering with a film potential-sensitive fluorochrome, the water surface fluctuations can be reduced and S/N can be improved by fitting this cap ② onto the top of the objective ① (XLFuor2X/340 or XLFluor4X/340).

5 OTHER OBSERVATION METHODS

5-1 Differential Interference Contrast Observation

★**The normal optical performance of DIC observation cannot be manifested if a plastic petri dish is used.**

◎DIC prisms (for revolving nosepiece and condenser), an analyzer and a polarizer are required for DIC observation. When the reflected light fluorescence illuminator is not used, the U-KPA intermediate tube is required to attach the analyzer.

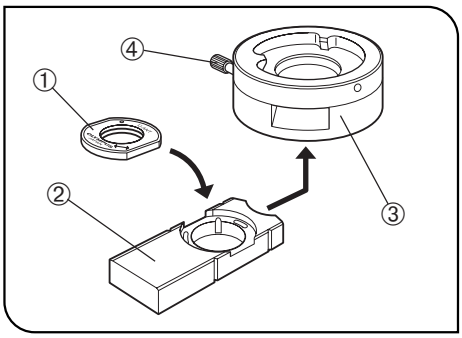


Fig. 28

1 Attaching the Analyzer (Fig. 28)

With the U-ANT Analyzer

◎Drop the U-ANT analyzer in the dummy slider of the U-KPA intermediate tube.

1. Place the U-ANT ① with the side with indications facing up, align the indices and drop the analyzer into the dummy slider ② (the analyzer will be absorbed by magnet).
2. Set the dummy slider ② back into the U-KPA ③ and tighten the clamping knob ④.

With the U-AN Analyzer

◎Insert the U-AN into the analyzer insertion slot of the reflected light fluorescence illuminator. (Refer also to the instruction manual of your reflected light fluorescence illuminator.)

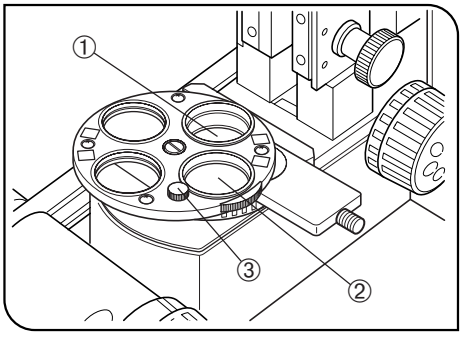


Fig. 29

2 Attaching the Polarizer (Fig. 29)

◎The performance of polarizer drops after it has been subjected to light for long hours. Replace it after about 500 hours of continuous use.
Drop the polarizer into the filter insertion position ① or ② with a push ring, and clamp with the push ring.

◎It is recommended to insert the polarizer in insertion position ①. This is because the polarizer rotation clamping knob ③ comes on the front of the microscope when the insertion position ① is engaged in the light path.

◎If you use the 32PO polarizer, the adjustment will be easier than with the 32POIR because the 32PO is brighter.
Also, the adjustment for IR reservation can be conducted under brighter light if you remove the IR filter (32BP775 or 32IR900) during adjustment.

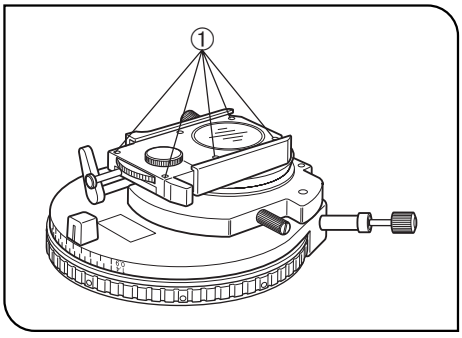


Fig. 30

When Using the U-UCD8

★**The polarizer built into the U-UCD8 is not necessary with this microscope. Remove it as described below.**

1. Using a Phillips precision screwdriver, remove the 6 clamping screws ① retaining the polarizer cover at the bottom side of the condenser.
2. Remove the cover to expose the polarizer and remove it together with the frame. (Retain the removed polarizer carefully for future use with a microscope other than the BX51WI.
3. Attach the polarizer cover to the original position.

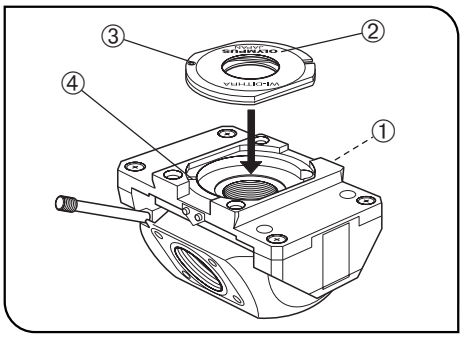


Fig. 31

3 Attaching the DIC Prisms (for Revolving Nosepiece) (Fig. 31)

◎The DIC prisms for use in the revolving nosepiece include the WI-DICTHRA2 (high-resolution type) and WI-DICT2 (middle-contrast type). The revolving nosepieces in which a DIC prism can be inserted are the WI-SRE3 and WI-SNPXLU2.

The DIC prisms (WI-DICT or WI-DICTHRA) cannot be attached to the revolving nosepieces (WI-SRE3 or WI-SNPXLU2) because of their shapes and sizes.

1. Remove the revolving nosepiece from the revolving arm.
   Sufficiently loosen the drop prevention screw ① with a Phillips precision screwdriver.
2. Hold a DIC prism ② with the side with indications facing up, and insert in by aligning the positioning pin ③ with the groove ④ on the revolving nosepiece.
   After the insertion, fully tighten the drop prevention screw ①.
3. Attach the revolving nosepiece onto the revolving arm.

4 Attaching the DIC Prisms (for Condenser) (Fig. 32-35)

◎DIC prisms can be inserted in three types of condensers including the WI-UCD, WI-DICD and U-UCD8*.

* Do not use the U-UCDTP530 one-wave plate for the U-UCD8 but use the exclusive WI-TP137 quarter-wave plate.

■List of DIC System Combinations

		Shearing Amount	
		Small (High resolution)	Medium (Middle contrast)
Condenser		WI-UCD WI-DICD	U-UCD8 (with WI-TP137)*
DIC prism (for revolving nosepiece)		WI-DICTHRA2	WI-DICT2
Objective		DIC prism (for condenser)	
Magnification	10X	U-LDPW10H (small)	U-DP10
	20X	U-LDPW20H (small)	———
	40X	U-LDPW40H (large)	U-DPO40S
	60X	U-LDPW60H (large)	U-DPO60S
	100X		U-DP100
	XLU20X	**U-LDPXLU20HR (large)	———
Application	CCD observation (Surface to deep)	Observation of relatively shallow area (0 to 100 μm)	Observation of relatively deep area (50 to 150 μm)
	Binocular observation (Surface layer only)	Optimum for surface layer observation.	Less suitable for surface layer observation than the high-resolution type.

** The actual view is equivalent to the middle-contrast type because of lower magnification and higher NA than usual.

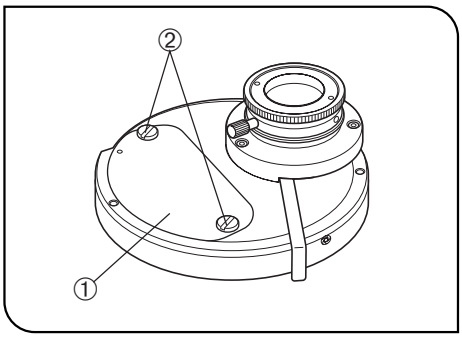


Fig. 32

With the WI-UCD Condenser (Figs. 32-34)

◎When selecting the brightfield (BF) light path using the WI-UCD, leave one DIC prism (large) mount position empty.

1. Remove the WI-UCD condenser from the microscope frame.
2. Remove the condenser cover ① by loosening the retaining screws ② using a coin, etc.

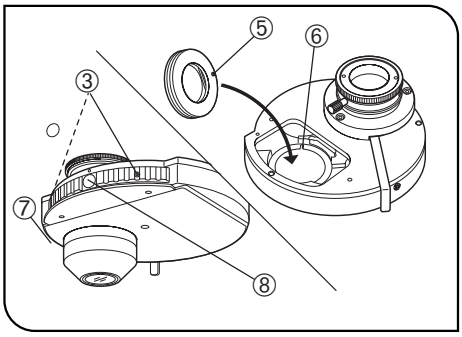


Fig. 33

3. Attach the suitable DIC prism for the objective in use as described below.
   - Using the knob provided with the condenser, loosen the two DIC prism clamping screws ③ until the rotatable limits.
   - Rotate the turret by 90° counterclockwise, and drop in the DIC prism by aligning its positioning pin ⑤ with the positioning groove ⑥ in the hole of the turret ④. (Fig. 33)

★Be careful not to touch the prism inside the frame.

A. Rotate the turret ④ by 90° clockwise (Fig. 34) and tighten the two DIC prism clamping screws ③ uniformly using the dedicated knob provided with the condenser. (Figs. 33 & 34)

★Do not tighten the screws too much, or the prism frame may be deformed.

B. Rotate the turret ④ by 90° clockwise (Fig. 34), and attach the index sticker ⑦ provided with the DIC prism onto the side ⑧ of the condenser turret ④ so that the index sticker is upside down. (Figs. 33 & 34)

4. After attaching all of the required DIC prisms, attach the cover ① and tighten the retaining screws ②. (Fig. 32)
5. Attach the condenser back onto the microscope frame.

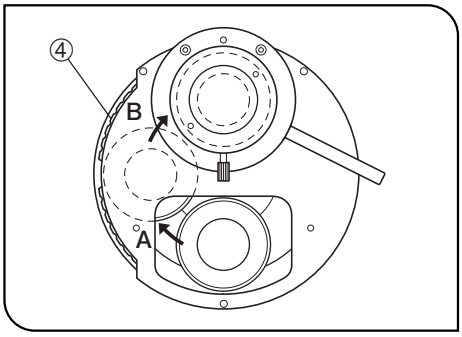


Fig. 34

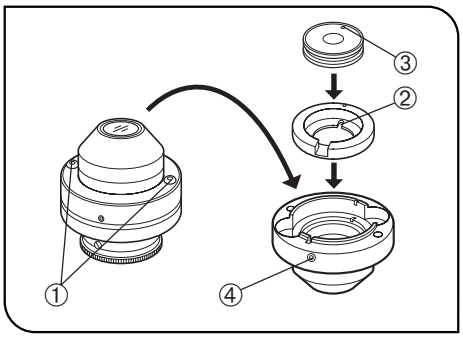


Fig. 35

With the WI-DICD Condenser (Fig. 35)

◎The WI-DICD should be attached after completing the polarizer position as described in item **5**.

1. Remove the WI-DICD condenser from the microscope frame.
2. Remove the two clamping screws ① using the Allen screwdriver provided with the microscope, then place the top of the condenser upside down.
3. When the DIC prism for use with the objective in use is a small DIC prism, drop it in by aligning the positioning groove ② on the adapter located on the inner side with the pin ③ of the prism.
   When the DIC prism for use with the objective is a large DIC prism, remove the adapter and drop in the DIC prism.

◎Retain the adapter for future possible use.

4. Tighten the clamping screws ④ with the knob provided with the condenser.
5. Attach the condenser on the microscope again.

With the U-UCD8

◎Attach the DIC prism by referring to the instruction manual provided with the U-UCD8.

5 Adjusting the Polarizer Position (except the U-UCD8) (Figs. 36-38)

★This adjustment is not necessary when the U-UCD8 is used. However, be sure to insert the WI-TP137 quarter-wave plate in a position where the U-UCDTP530 one-wave plate for the U-UCD8 is otherwise inserted.

◎This adjustment is possible without removing the DIC prism (for revolving nosepiece). However, it is not possible if a DIC prism for condenser is engaged in the light path. Remove or disengage the DIC prism for condenser as described below.

- WI-DICD: Remove the DIC prism.
- WI-UCD: Rotate the turret to engage a position without DIC prism.

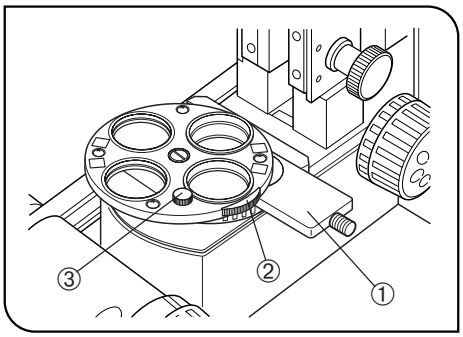


Fig. 36

When the U-LH100IR Lamp Housing Is Used

⚠Be sure to take the following measure to protect your eyes from the IR rays.

◎Insert the IR cut filter (light blue) provided with the microscope into the filter slider ①, then push it in to engage it. (See page 10.)

1. Remove the condenser from the microscope.
2. Remove an objective and engage the position without the objective in the light path.
3. Engage the polarizer and analyzer in the light path (page 19) and turn the transmitted light on.

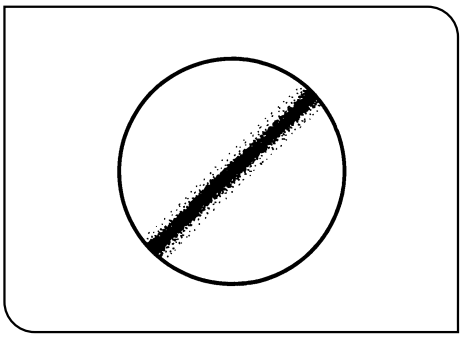


Fig. 37

4. Remove the eyepiece from the eyepiece sleeve, look into the sleeve, turn the polarizer rotation dial ② so that the black interference stripe (Fig. 37) is darkest, and tighten the clamping knob ③.
5. Engage an objective (as low-magnification as possible) in the light path, attach the condenser and bring the specimen surface into focus.

Important **The interference stripe is less clearly visible when the specimen is thick. In this case, it is recommended to bring a scratch or like on the bottom of the petri dish to facilitate the subsequent adjustment operation.**

◎With the WI-DICD, do not attach the DIC prism.
With the WI-UCD engage a position without DIC prism in the light path.

6. If the condenser has not been centered yet, center it (page 16).

◎The interference stripe is not visible clearly if the field iris is focused insufficiently.

7. Turn the quarter-wave plate rotation ring ① so that the black interference stripe seen at the center of the eyepiece sleeve's field of view, then tighten the clamping knob ②. Ignore the short interference stripes in the surroundings in this adjustment.
Since this adjustment renders the field of view dark, observation cannot be started unless the observation method described in the next item is employed.
Now the adjustment is complete.

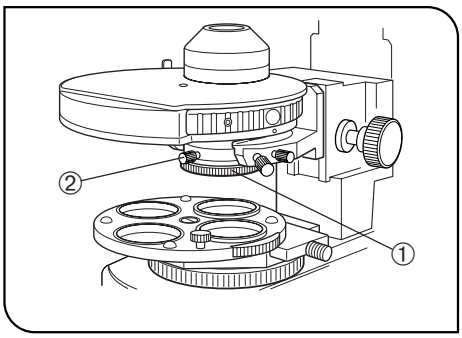


Fig. 38

- Attach the eyepiece and objective again to the microscope frame.
- With the WI-DICD, remove it and mount the required DIC prism.
- When an IR cut filter is used, remove it and mount the required filter.

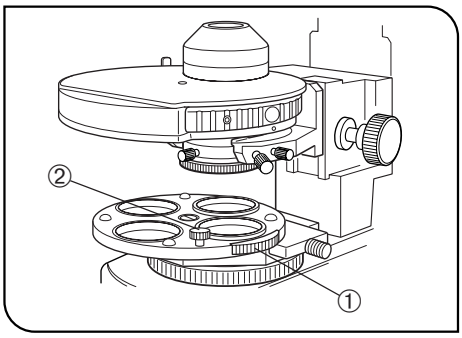


Fig. 39

6 Observation Method (Fig. 39)

1. Engage the objective to be used in the light path.
2. When the WI-UCD or U-UCD8 condenser is used, engage the DIC prism matching the objective in the light path by rotating the turret.
3. Place the specimen on the stage and bring the specimen into focus.

◎The contrast may be improved by stopping down the aperture iris diaphragm to an optimum aperture.

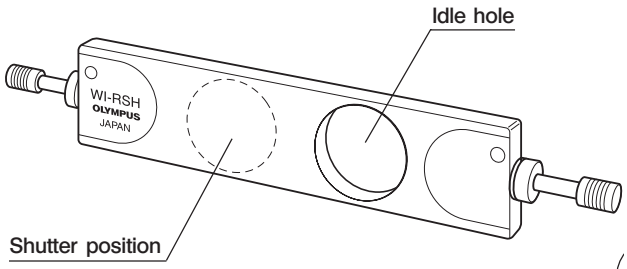
4. Rotate the polarizer dial ① on the filter turret to obtain optimum contrast for the specimen. Tighten the clamping knob ② if required.

5-2 Reflected Light Fluorescence Observation

◎Refer to the instruction manual of your reflected light fluorescence system.

Illuminator Shutter WI-RSH

◎The shock during observation can be reduced by using this optional shutter in place of using the shutter built into the BX-URA2 or BX-RFA reflected light illuminator.



(To be used by inserting in the 6-position filter/polarizer insertion slot of the reflected light illuminator.)

5-3 Infrared Light (IR)/Differential Interference Contrast (DIC) Observation

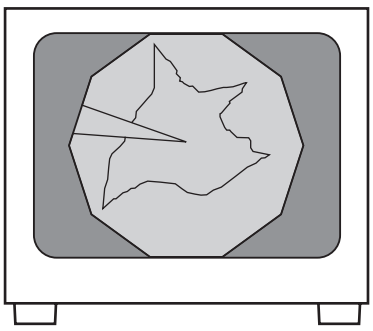
◎The IR rays (775 or 900 nm) transmit the specimen by about 4 or 5 times more than visible light (550 nm). Therefore, the IR observation is suitable for observing deep areas of a thick brain slice or optic nerve specimen.

1 Introduction

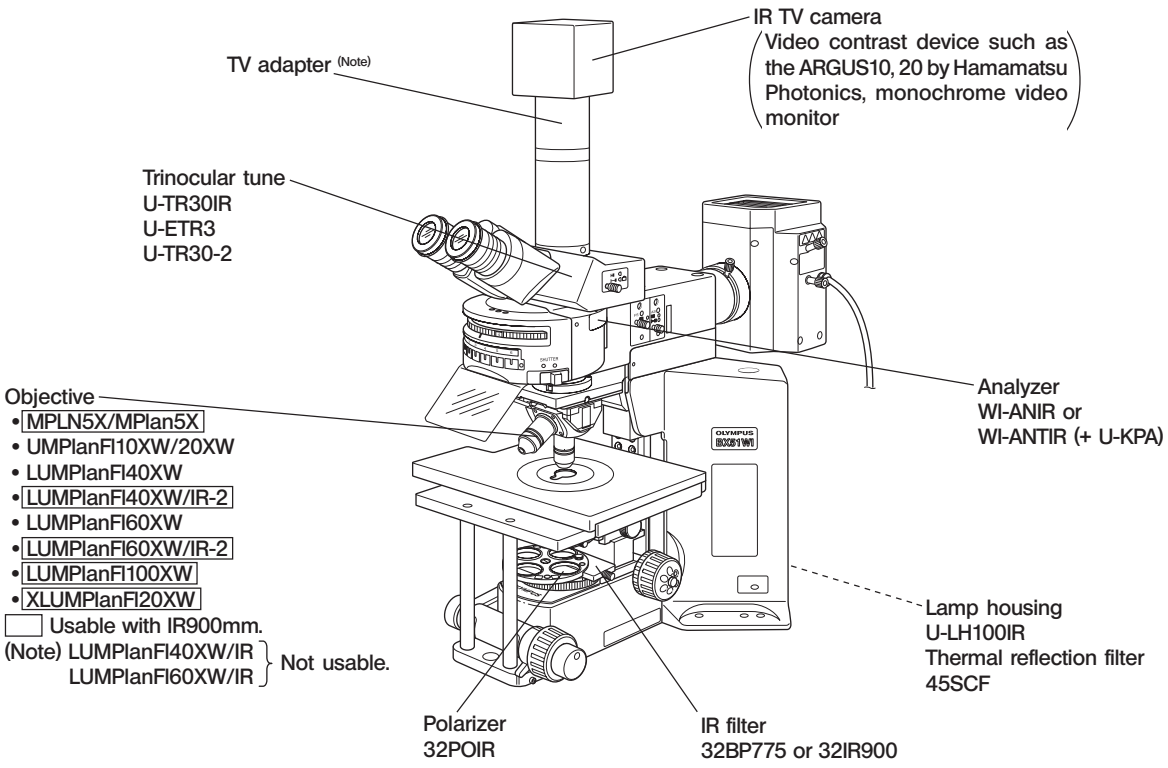
1. Since the IR wavelength used is 775 to 900 nm, the TV camera in use should be sensitive in the wavelength used (Example: C2741-79 CCD camera mfd. by Hamamatsu Photonics).

▲The IR light is harmful to your eyes. Avoid visual observation and use the TV monitor whenever possible. Should visual observation be used, mount the IR cut filter (light blue) provided with the filter turret and engage the IR cut filter in the light path. (See pages 10 and 22.)

2. To reduce the influence of heat on the specimen, stop down the field iris diaphragm of the BX51WI microscope as small as possible. However, note also that the contrast may sometimes be improved by circumscribing the field iris image with the field of view.



3. To enable IR observation, the following modules should be replaced with those based on the IR specifications.

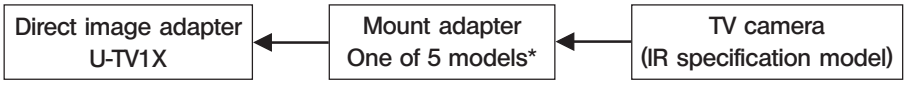


(Note) Notes for combination of the TV adapter, intermediate attachment and observation tube

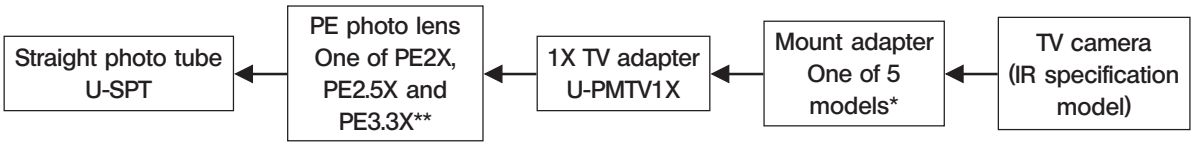
When an observation tube other than the U-TR30IR is used, select the TV adapter by referring to combinations a) to d) below.

★ **With IR observation, the combination with the U-PMTVC or U-DPT cannot manifest full performance.**

a) Combination for observing a wide field (direct image 1X)



b) Combination for magnified observation of objective image (2X, 2.5X or 3.3X) ** The PE4X and PE5X cannot be used.



* The mount adapter should be one of: 1) U-CMAD3 mount adapter; 2) U-BMAD bayonet mount adapter; 3) U-IMAD Ikegami camera mount adapter; 4) U-SMAD Sony camera mount adapter; 5) combination of U-FMT F/T mount + U-TMAD T-mount adapter.

Note) When the contrast is enhanced rather excessively by an image processor, the central area of the monitored image may be made bright and noticeable.

c) Combination using C-mount adapter IR system (visible light to 1000 nm)

C-mount adapter IR system	←	C-mount TV camera
U-TVCAC U-PMTVC2XIR U-PMTVC4XIR		(IR specification model)

d) Combination using U-CA or U-ECA intermediate tube

One of these intermediate tubes can be used only in combination with the U-ETR3 or U-TR30IR trinocular tube. The TV adapter used in this combination should be one of that used in a), b) or c).

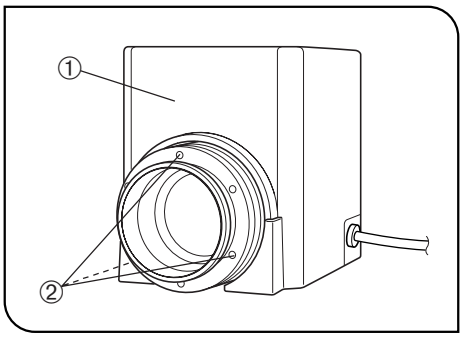


Fig. 40

2 Attaching the IR Modules (Figs. 40 & 41)

Thermal Reflection Filter 45SCF

1. Remove the collector lens of the U-LH100IR lamp housing ① by loosening the 3 clamping screws ② with an Allen wrench (width across flats of 2.5 mm).

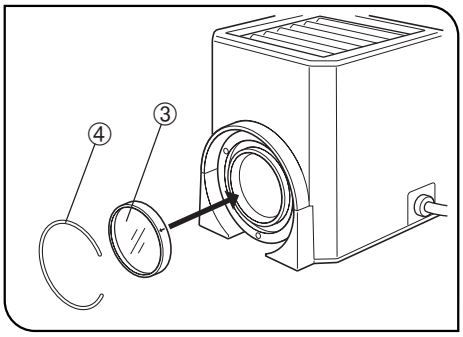


Fig. 41

2. While positioning the 45SCF filter ③ so that the arrow on its frame points in the opposite direction of the lamp housing, insert the filter in the lamp housing, and clamp by tightening the ring spring ④ provided with the filter.
3. Attach the collector lens to the original position.

IR Filter 32BP775 or 32IR900

◎ Be sure to insert the 32BP775 or 32IR900 IR filter in the filter slider below the filter turret. (For the mounting method, see page 10.)

★ **If the IR filter is inserted above the polarizer in the filter turret, the polarizer will be burnt.**

Other IR Modules

Also replace other required modules with the IR modules (see page 25).

3 DIC Observation Using IR

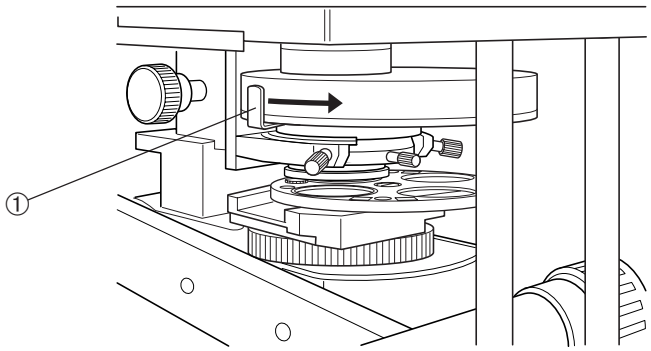
▲Since IR light is harmful to your eyes, use the monitor observation whenever possible even in adjustments.

1. First, perform adjustments for DIC observation without using IR.

◎Do not mount the IR filter and see Section 5-1, "Differential Interference Contrast Observation" on page 19.

Important

- **Focus the field iris diaphragm image (page 16).**
  Be sure to perform this adjustment accurately because it determines the visual performance using the IR light.
- **With DIC observation using IR, do not stop down the aperture iris diaphragm lever ① but leave the diaphragm open.**
  As the contrast can be enhanced using the video enhancement function of the CCD controller, the diaphragm should be left open in this adjustment so that the optical performance can be manifested fully.



2. Then engage the IR filter (32BP775 to 32IR900) in the light path by pushing in the filter slider.
3. While observing the monitor, perform DIC observation using IR.
   a) Turn the condenser turret to select the DIC prism matching the objective to be used (except the WI-DICD).
   b) Turn the revolving nosepiece to engage the objective to be used in the light path.

★Penetration of air bubbles inside the front lens of objective will deteriorate the view. To prevent this by removing the bubbles, turn the revolving nosepiece slightly to move the immersed objective to the left and right for a few times.

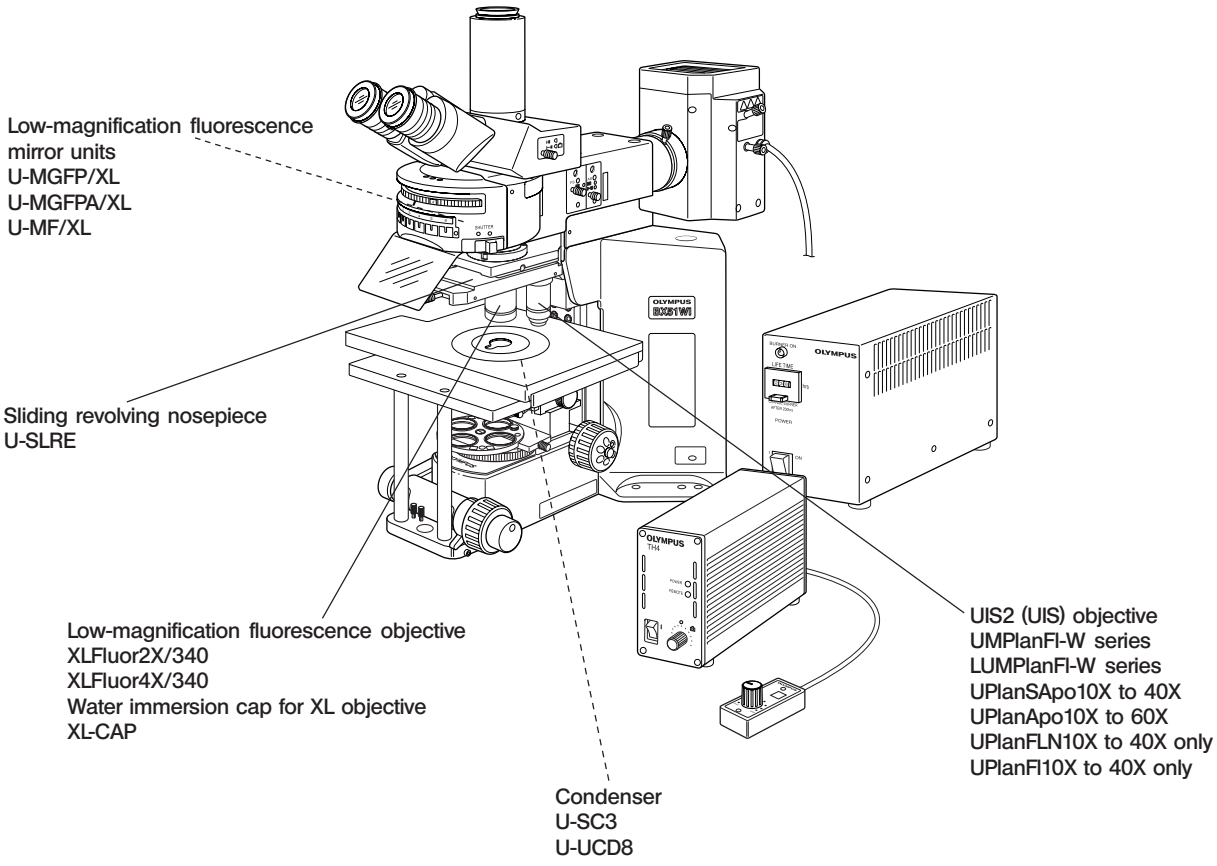
   c) Bring the specimen into focus by moving the objective up and down using the coarse and fine adjustment knobs.
4. Turning the 32POIR polarizer varies the density of the background. Set the polarizer to obtain optimum contrast for the specimen.

5-4 Macro Reflected Light Fluorescence Observation

◎The macro reflected light fluorescence observation makes possible bright, low-magnification fluorescence observation by combining low-magnification fluorescence mirror units and fluorescence objective.

1 Introduction

1. For the low-magnification fluorescence observation, use a low-magnification fluorescence mirror units.
   The increased observation beam diameter of the fluorescence mirror units brightens the fluorescence by about 25%.
   However, due to the large size of the fluorescence mirror units, they can be mounted only in every other position when the BX-URA2 or BX-RFA reflected fluorescence illuminator is used (a total of 3 units can be mounted on each illuminator).
2. When performing transmitted light brightfield observation using a low-magnification fluorescence objective (2X or 4X), also use the U-SC3 or U-UCD8 swinging condenser. If other condenser is combined, it will not be possible to illuminate the entire field of view.
3. During low-magnification fluorescence observation, objective switching or stage movement, be careful so that the UIS objective does not interfere with the specimen or culture container.
4. The low-magnification fluorescence objectives have been designed to manifest performances with no-covered dry specimens to specimens located 5 mm below water surface level.
   As a result, with water immersed specimens, the focused positions of these objectives are different from UIS objectives.
5. To enable macro reflected light fluorescence observation, the following modules should be replaced.



2 Attaching the Modules (Fig. 42 & 43)

Low-Magnification Fluorescence Mirror Units

◎Select the suitable mirror units for purpose of observation by referring to page 30.

◎If you want to fabricate optional mirror units, see page 30.

• Mount the mirror units as indicated in the instruction manual of your reflected light fluorescence system.
Note that mirror units can be mounted only in every other positions.

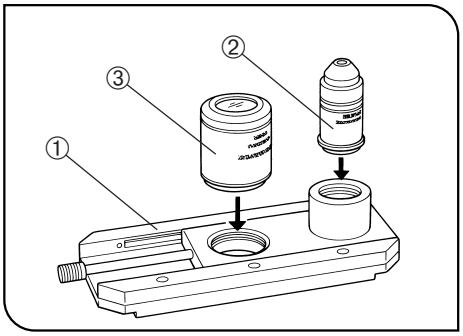


Fig. 42

Objective

1. Screw a UIS objective ② into the position on the deeper side of the U-SLRE sliding revolving nosepiece ①.
2. Screw a XLFluor2X/340 or XLFluor4X/340 low-magnification fluorescence objective ③ into the position on the shallower side of the U-SLRE.

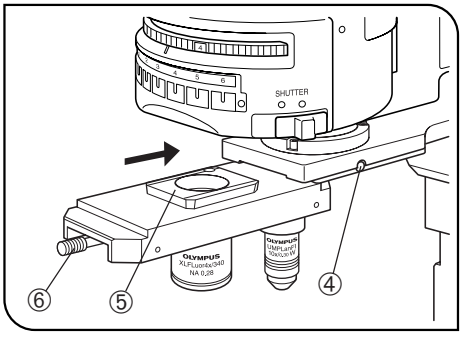


Fig. 43

Sliding Revolving Nosepiece U-SLRE

1. Raise the revolving nosepiece mount fully by rotating the coarse adjustment knob of the microscope frame.
2. Loosen the revolving nosepiece mount screw ④ on the microscope frame using the Allen screwdriver provided with it.
3. Align the mount dovetail ⑤ of the sliding revolving nosepiece with the revolving nosepiece mount dovetail and gently slide the sliding revolving nosepiece all the way in from the front as shown in the figure.
4. Clamp the revolving nosepiece by tightening the revolving nosepiece mount screw ④.

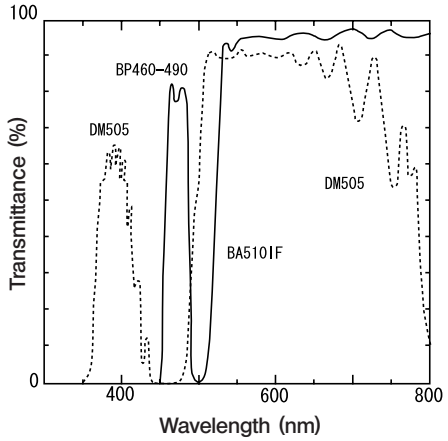
Swinging Condenser U-SC3/U-UCD8

★Attach the U-SC3 with the top lens swung out.
The condenser top lens should be swung out when using the 2X or 4X objective.

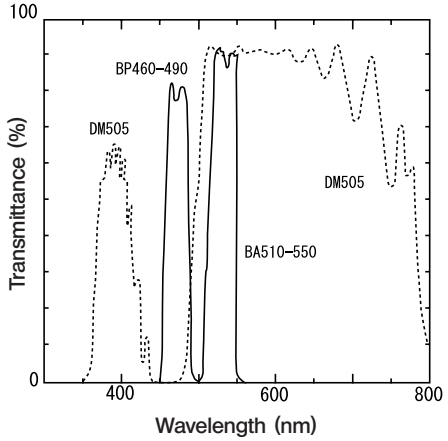
3 Filter Characteristics of Fluorescence Mirror Units

Excitation Method	Mirror Unit	Dichroic Mirror	Excitation Filter	Barrier Filter	Application
IB	U-MGFP/XL	DM505	BP460-490	BA510IF	For EGFP, S65T, RSGFP. (U-MGFPA/XL is for fluorochrome separation.)
	U-MGFPA/XL			BA510-550	

1. IB excitation (wide bandwidth) U-MGFP/XL



2. IB excitation (wide bandwidth) U-MGFPA/XL

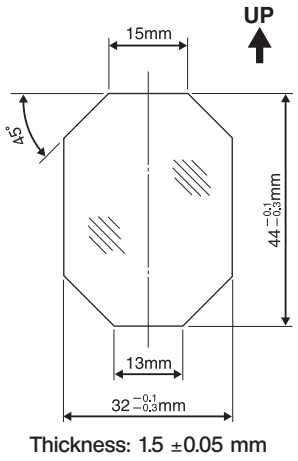


4 Fabricating Optional Mirror Unit

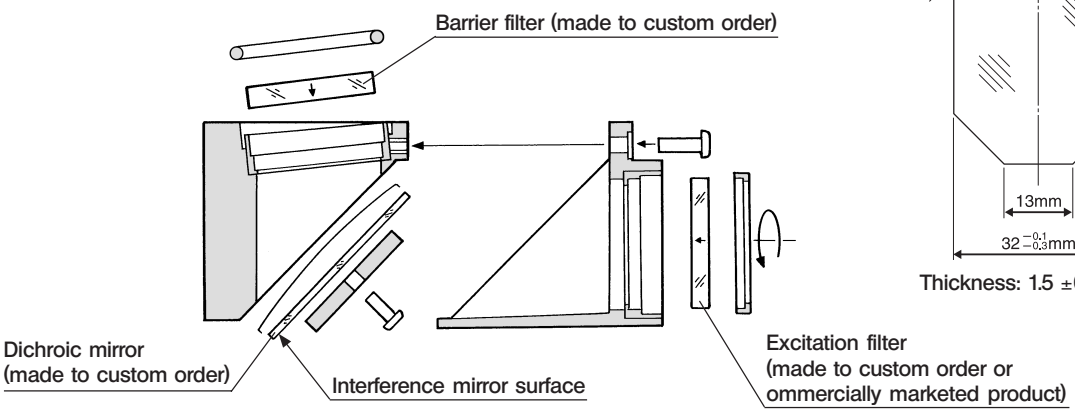
◎An optional mirror unit can be fabricated by attaching the custom-order barrier filter, excitation filter and dichroic mirror to the U-MF/XL.

Dimension Conditions of Optical Components of Mirror Unit

- Barrier filter: $\phi 32^{-0.1/-0.2}$ mm, max. thickness 4 mm
- Excitation filter: $\phi 25^{-0.1/-0.2}$ mm, max. thickness 6 mm
- Dichroic mirror: See figure on the right.



Thickness: 1.5 ±0.05 mm



★When replacing the dichroic mirror, take special care not to stain it by leaving fingerprints, etc.

6 TROUBLESHOOTING GUIDE

Under certain conditions, performance of the microscope may be adversely affected by factors other than defects. If problems occur, please review the following list and take remedial action as needed.
If you cannot solve the problem after checking the entire list, please contact your local Olympus representative for assistance.

Problem	Cause	Remedy	Page
1. Optical System			
a) The bulb does not light.	The bulb is burned out.	Replace the bulb.	41
b) The bulb lights but the field of view is dark.	The aperture or field iris diaphragm is opened in sufficiently.	Open the aperture and field iris diaphragms.	16
	The condenser is in too low a position.	Adjust the condenser height.	16
	The light path selector knob of is set to position 📷.	Set the light path selector knob to position 👁📷 or 👁.	15
	The voltage selector knob is set to a low voltage position.	Set it to the high voltage position.	10
c) Field of view is obscured or not evenly illuminated.	The light path selector knob is in an intermediate position.	Set the light path selector knob to a click position according to the purpose.	15
	The revolving nosepiece is not in a click position.	Set it in a click position.	13
	The revolving nosepiece is installed incorrectly.	Secure it by pushing in the sliding dovetail all the way until the stopper.	38
	The filter turret or filter slider is incorrectly engaged in the light path.	Engage them correctly in the light path.	10
	The condenser is not centered.	Adjust the centering.	16
	The frost switching lever is set to an intermediate position or OUT.	Engage the frost filter correctly in the light path.	11
	The field iris diaphragm is closed too much.	Open it sufficiently.	16
	The lamp bulb is not installed correctly.	Push the halogen bulb terminals all the way into stop position.	41
d) Dirt or dust is visible in the field of view.	Dirt/dust on eyepiece.	Clean thoroughly.	2
	Dirt/dust on condenser top lens.		
	Dirt/dust on specimen.		
e) Image glares.	The condenser is set to too low a position.	Adjust the condenser height.	16
	The aperture iris diaphragm is closed too much.	Open it sufficiently.	17
f) Visibility of observed image is poor. • Image is not sharp. • Contrast is poor. • Details are poorly visible.	The objective in use is not designed for UIS2 (UIS) series.	Replace with a specified objective for UIS2 (UIS) series.	35
	The revolving nosepiece is installed incorrectly.	Secure it by pushing in the sliding dovetail all the way until the stopper.	38
	The objective is engaged incorrectly in the light path.	Make sure that revolving nosepiece clicks into place correctly.	13
	Air in the objective front lens.	Remove the air.	–

Problem	Cause	Remedy	Page
f) Visibility of observed image is poor. • Image is not sharp. • Contrast is poor. • Details are poorly visible.	The specimen such as a brain slice is fixed poorly.	Fix it correctly.	–
	Bubbles attached to the objective front lens.	Remove the bubbles.	–
	Too small quantity of solution in the petri dish.	Supply sufficient solution in the petri dish.	–
	The petri dish is tilted.	Place the petri dish correctly on the stage.	12
	Dirt/dust on the objective front lens.	Clean it thoroughly using neutral detergent.	–
	Dust/dirt on the condenser.	Clean it thoroughly.	2
g) One side of image is blurred.	The revolving nosepiece is installed incorrectly.	Secure it by pushing in the sliding dovetail all the way until the stopper.	38
	The objective is engaged incorrectly in the light path.	Make sure that revolving nosepiece clicks into place correctly.	13
	The objective is placed incorrectly (may be loose) in the revolving nosepiece position.	Insert the objective all the way into the revolving nosepiece position until it is stopped.	–
	The stage center plate is tilted.	Correct the tilt.	–
h) Image appears to waver.	The revolving nosepiece is installed incorrectly.	Secure it by pushing in the sliding dovetail all the way until the stopper.	38
	The objective is engaged incorrectly in the light path.	Make sure that revolving nosepiece clicks into place correctly.	13
	The objective is placed incorrectly (may be loose) in the revolving nosepiece position.	Insert the objective all the way into the revolving nosepiece position until it is stopped.	–
	The condenser is centered incorrectly.	Center it correctly.	16
i) The field of view becomes brighter only slightly although the voltage is increased.	The condenser is centered incorrectly.	Center it correctly.	16
	The condenser is in too low a position.	Adjust the condenser height.	16
2. Electrical System			
a) The bulb intermittently lights and goes out.	The bulb is nearly burnt out.	Replace the bulb.	41
	A cord or connector is not properly connected.	Connect cords and plugs securely.	–
b) The lamp bulb burns out soon after lighting.	The bulb in use is not the specified lamp.	Replace with a standard bulb.	41
c) The brightness cannot be varied with the light intensity control.	No lamp bulb is installed.	Attach a lamp bulb.	41
	The lamp bulb is burnt out.	Replace the lamp bulb.	41
	The lamp housing output connector is unplugged.	Plug the lamp housing output connector.	–

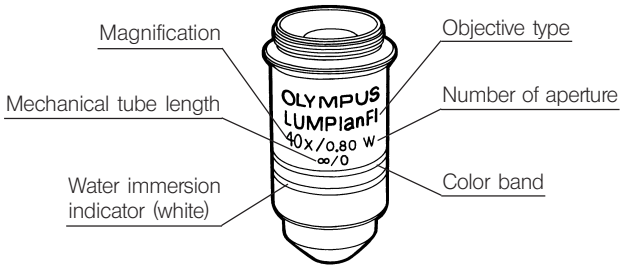
Problem	Cause	Remedy	Page
3. Coarse/Fine Adjustment Knobs			
a) Coarse adjustment knob is too heavy to rotate.	The rotation tension adjustment ring is secured tightly.	Fully loosen the ring by turning it counterclockwise.	11
	The pre-focusing lever is locked.	Release the pre-focusing lever.	11
b) The objective cannot be lowered enough.	The pre-focusing lever is functioning.	Release the pre-focusing lever.	11
c) Objective confocality is not achieved with the WI-SRE3.	Adjustment is not correct.	Adjust correctly.	38,39
4. Observation Tube			
a) The field of view of one eye does not match that of the other.	The interpupillary distance is incorrect.	Adjust interpupillary distance.	14
	Incorrect diopter adjustment.	Adjust diopter.	14
	Different eyepieces are used on the left and right.	Change one eyepiece to match the other so that both sides are of the same type.	–
	You are not accustomed to parallel optical axis.	When looking into eyepieces, do not stare at image from the beginning but see the overall field of view. It is sometimes recommended to turn your eyes away from eyepieces, look far off and look into eyepieces again.	–
5. Stage			
a) Stage travel in the horizontal (X-axis) direction stops in the middle.	The specimen is set incorrectly.	Place the specimen correctly.	12
b) The X-axis and/or Y-axis stage knobs are too light or too heavy to rotate.	The X-axis and/or Y-axis rotation tension is not adjusted properly.	Adjust the knobs to optimum tension.	12

7 SPECIFICATIONS

Item	Specification			
1. Optical system	UIS2 (UIS) (Universal Infinity System) optical system (Infinity correction)			
2. Illumination system	Transmitted Kohler illumination built in (FN 22) 12 V, 100 W long-life halogen bulb (pre-centered) Average life time: Approximately 2000 hr. when used as directed. Light intensity voltage range: 2.5 ∿ 12.6 V DC (continuously variable), 8.4 A max. Power consumption: 140 W			
3. Focusing system	Revolving nosepiece height movement by roller guide (rack & pinion) Stroke per rotation: 0.1 mm (fine), 15 mm (coarse) Full stroke range: 25 mm Pre-focusing lever Coarse adjustment knob: Tension adjustment possible			
4. Revolving nosepiece	Model	WI-SRE3 Swinging Revolving Nosepiece	U-SLRE Sliding Revolving Nosepiece	WI-SNPXLU2 Single-Position Revolving Nosepiece
	Attachable modules	DIC prisms mountable	———	DIC prisms mountable
5. Observation tube	Model	U-TR30-2 Widefield Trinocular	U-ETR3 Widefield, Upright-Image Trinocular	
	Field number	22		
	Tube tilting	30°	25°	
	Interpupillary distance adjustment	50 mm to 76 mm		
	Light path selection	3-step switching: ①Binocular 100% ②Binocular 80% TV/photo 20% ③TV/photo 100%	2-step switching: ①Binocular 100% ②TV/photo 100%	
6. Stage	Model	IX-SVL2		
		Stage with Bottom-Side Knobs		
	X/Y movement mechanism	X-axis/Y-axis knob tension adjustable Movement range: 50 mm (X-axis) x 45 mm (Y-axis)		
7. Long-WD condenser	Model	WI-UCD Universal Condenser	WI-DICD DIC Condenser	WI-OBCD Oblique Condenser
	N.A.	0.8		
	Working distance	5.7 mm		
	Aperture iris	Variable aperture iris diaphragm		
	Turret	4-position	———	
	DIC prisms	Max. 4 prisms can be mounted.	Only 1 prism can be mounted.	———
	Other	Quarter-wave plate built in		Variable oblique iris built in
8. Operating environment	• Indoor use • Altitude: Max. 2000 m • Ambient temperature: 10° to 40°C (50° to 104°F) • Maximum relative humidity: 80% for temperatures up to 31°C (88°F), decreasing linearly through 70% at 34°C (93°F), 60% at 37°C (99°F), to 50% relative humidity at 40°C (104°F) • Supply voltage fluctuations: ±10% • Pollution degree: 2 (in accordance with IEC60664) • Installation (overvoltage) category: II (in accordance with IEC60664)			

8 OPTICAL CHARACTERISTICS

The following table shows the optical characteristics of combinations of eyepieces and objectives. The figure on the right shows the performance data engraved on the objectives.



NOTE

Refer to the latest catalogue or consult your local Olympus representative for the updated information on the eyepieces and objectives that can be combined with this microscope.

Optical character / Objective	Power	N.A.	W.D. (mm)	Resolution (μm)	Eyepiece WHN10X (FN22) Total Power	Eyepiece WHN10X (FN22) Focal Depth (μm)	Eyepiece WHN10X (FN22) Actual Field	Remark
MPLN Plan Achromat (FN 22)	5X	0.1	20.0	3.36	50X	98	φ4.4	Water immersion impossible
UMPlanFI-W Water Immersion Universal Plan Semi-Apochromat (FN 26.5)	10XW 20XW	0.30 0.50	3.30 3.30	1.10 0.67	100X 200X	20 6.1	φ2.2 φ1.1	
LUMPlan-W LUMPlan-W/IR-2 Long-WD Water Immersion Universal Plan Semi-Apochromat (IR-2: For infrared light) (FN 26.5)	40XW 40XW/IR-2 60XW 60XW/IR-2 100XW	0.80 0.80 0.90 0.90 1.00	3.30 3.40 2.00 2.00 1.50	0.42 0.42 0.37 0.37 0.34	400X 400X 600X 600X 1000X	2.0 2.0 1.3 1.3 0.83	φ0.55 φ0.55 φ0.37 φ0.37 φ0.22	
XLUMPlanFI-W High-NA Water Immersion (FN 22)	20XW	0.95	2.00	0.35	200X	3.20	φ1.1	
XLFluor Low-Power Fluorescence (FN 22)	2X/340 4X/340	0.14 0.28	20.0 28.4	2.40 1.20	20X 40X	132 33.0	φ11 φ5.5	Water immersion impossible Water immersion impossible

★ After using a water immersed objective, be sure to clean the extremity using neutral detergent. If the extremity is left without cleaning, contamination remains and the objective performance will deteriorate.

9 ASSEMBLY

9-1 Assembly Diagram

The diagram below shows the sequence of assembly of the modules. The numbers indicate the order of assembly.
The module numbers shown in the following diagram are merely the typical examples. For the modules with which the module numbers are not given, please consult your Olympus representative or the latest catalogues.

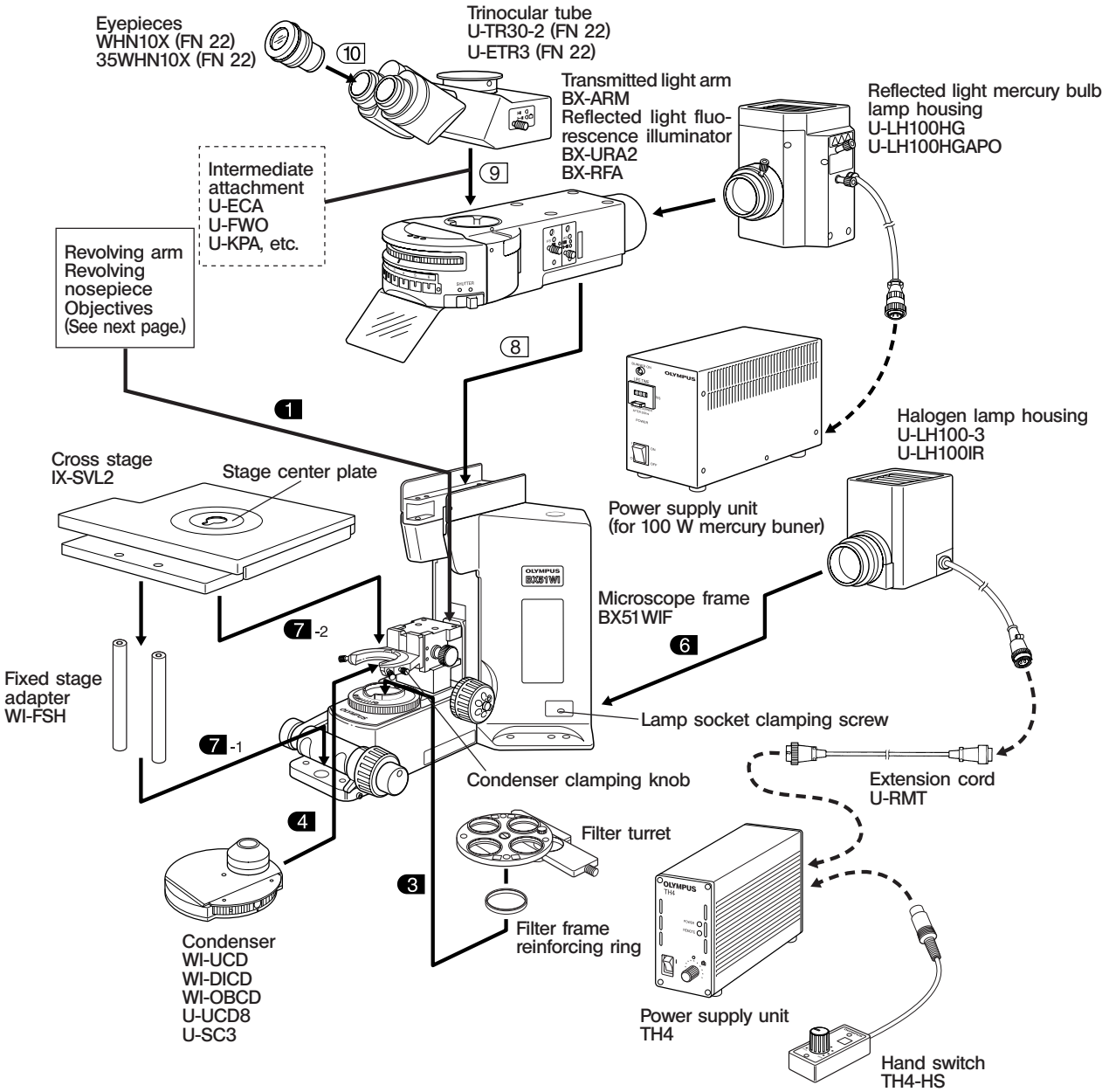
★ **When assembling the microscope, make sure that all parts are free of dust and dirt, and avoid scratching any parts or touching glass surfaces.**

Assembly steps enclosed in ■ will be detailed on the subsequent pages.

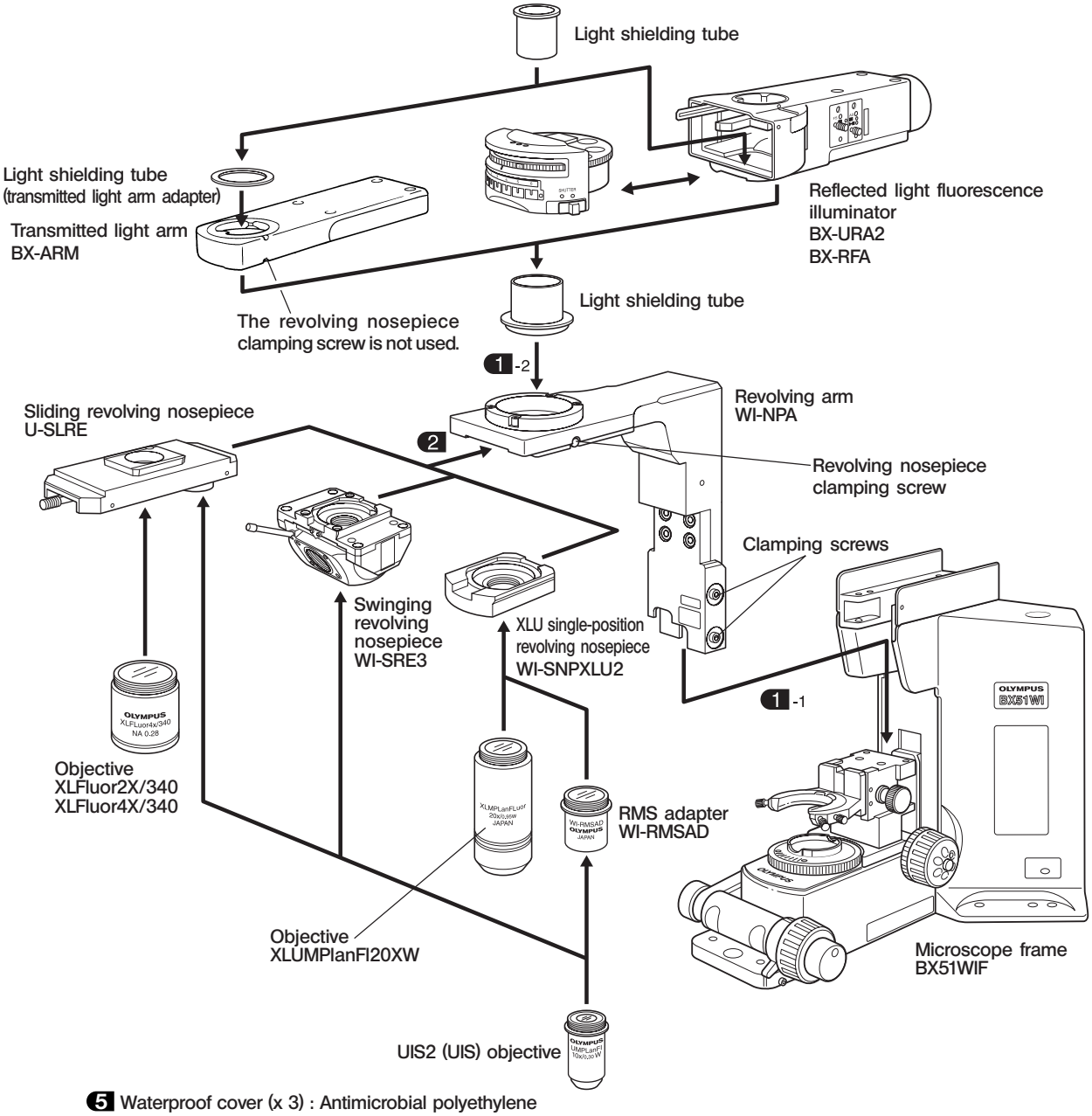
◎ All assembly operations are possible by using the Allen screwdriver ( ) provided with the microscope frame. However, the reflected light illuminator or transmitted light arm should be attached using the Allen wrench provided with the illuminator or arm to tighten the clamping screws. (To assure the performance, please have your dealer assemble the illuminator or arm.)

The filter turret and cross stage are to be cleaned respectively using the special tools provided with them.

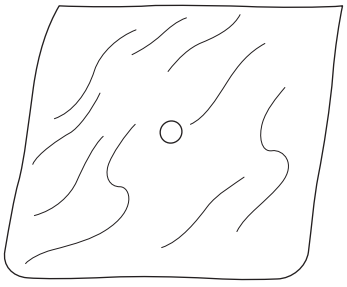
(Note) For the detailed assembly procedures of the reflected light fluorescence system and the TH4 power supply unit, refer to their instruction manuals.



Revolving Arm, Revolving Nosepiece, Objectives



5 Waterproof cover (x 3) : Antimicrobial polyethylene



Magnets

Magnet support plates
(with double-side adhesive tape)

Clamping band

9-2 Detailed Assembly Procedures

◎When performing observation on a desktop, attach the provided rubber feet (x 4) to the front (x 2) and rear (x 2) parts on the bottom of the base.

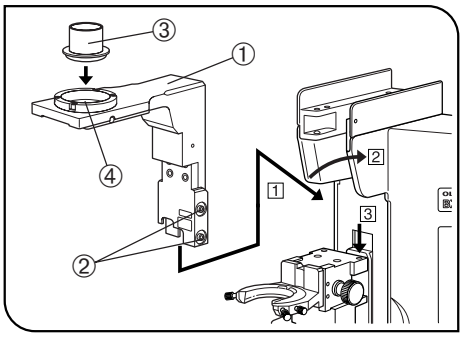


Fig. 44

1 Attaching the Revolving Arm (Fig. 44)

1. Loosen the 2 clamping screws ② of the revolving arm ① using an Allen screwdriver and fit the arm into the mount dovetail on the microscope frame along the direction of arrow from 1 to 3.
2. Push the revolving arm all the way until it is stopped, then tighten the clamping screws.
3. Screw in the light shielding tube ③ into the retaining screw ④.

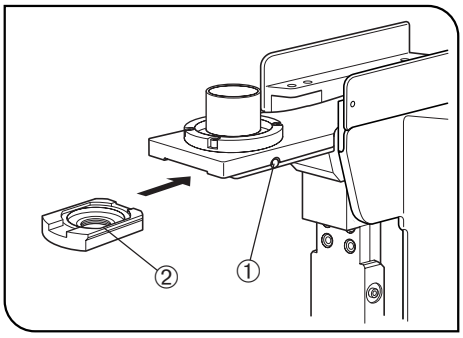


Fig. 45

2 Attaching the Revolving Nosepiece (Fig. 45)

◎The attaching procedure is common for the U-SLRE, WI-SRE3 and WI-SNPXLU2 revolving nosepieces, except that the WI-SNPXLU2 should be inserted so that the round end comes on the front.

CAUTION **In DIC observation using the WI-SRE3 or WI-SNPXLU2 revolving nosepiece, attach the DIC prisms before attaching the revolving nosepiece (see page 20).**

1. Loosen the revolving nosepiece clamping screw ① using an Allen screwdriver, and slide in the revolving nosepiece ② along the mount dovetail.
2. Push the revolving nosepiece all the way in and tighten the clamping screw.

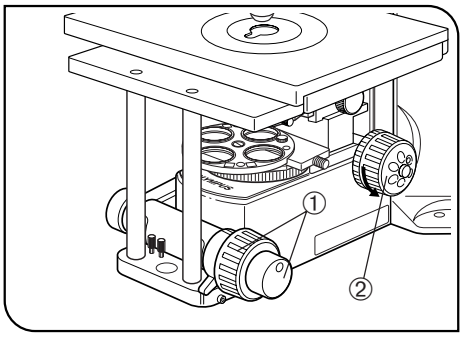


Fig. 46

WI-SRE3 Only

◎When the microscope has been assembled, the confocality adjustment and centering of the two objectives can be performed.

Adjusting the Confocality of Objective (Figs. 46 & 47)

◎To maintain the accurate focusing even after the objective is switched, the height of the objective with the higher focal point is corrected by attaching a washer.
Nine washers with three kinds of thickness (10, 30 and 50 μm), three per kind, are provided with the microscope.

1. Engage the objective on the front side in the light path. Use the coarse/fine adjustment knobs on the front side to adjust focusing.

◎Accurate focus cannot be achieved if the coarse/fine adjustment knobs on the back side are used.

2. To find the confocality difference, read the scale indication of the fine adjustment dial ② (1 scale graduation: 1 μm).

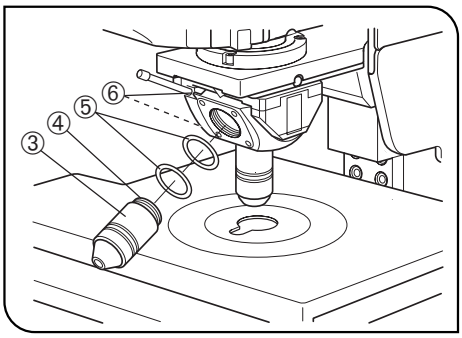


Fig. 47

3. Engage the objective on the back side in the light path.
   Adjust focusing, read the scale indication of the fine adjustment knob and obtain the difference from the value read in step 2 above.
4. The objective to use the washer is determined by the rotation direction of the fine adjustment dial ②. (Fig. 46)
   • Direction of the arrow: Objective on the back side.
   • Opposite direction to the arrow: Objective on the front side.
5. Remove the objective ③, fit the washer ⑤ with the required thickness on the screw ④, then attach the objective again.
6. Switch the objective and confirm that confocality is implemented.

★ **The confocality adjustment may sometimes be unable to implement perfect confocality.**

Centering the Objective (Fig. 47)

◎The centering mechanism is provided only for the objective on the front side.

1. Adjust focusing using the objective on the back side, then move the target region in the specimen on the center of the field.
2. Switch the objective to that on the front side.
3. Insert the centering knobs into the threaded holes ⑥ and turn them to move the target in the objective on the center.

◎After completing centering, store the centering knobs in the accommodation position on the front side of the microscope (page 4) so as not to lose them.

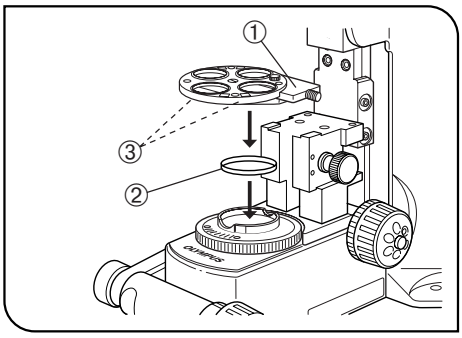


Fig. 48

3 Attaching the Filter Turret (Fig. 48)

◎When a bridge stage or a stage with similar design is used, the filter slider ① can be attached so that it faces toward the front of the microscope.

1. Drop in the filter frame reinforcing ring ② into the filter holder on the microscope base.
2. Loosen the 3 filter turret clamping screws ③ using the provided Allen wrench.
3. Fit the filter turret on the filter holder and tighten the clamping screws ③.
4. For the insertion of the polarizer and filters, see page 10.

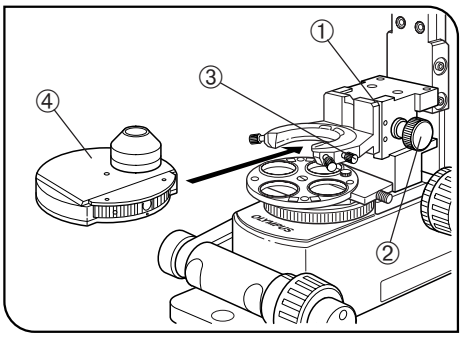


Fig. 49

4 Attaching the Condenser (Fig. 49)

★**When attaching a condenser other than the WI-UCD, remove the upper limit stopper screw ① of the condenser holder with an Allen screwdriver.**

◎In DIC observation, attach the DIC prism (for condenser) before attaching the condenser onto the microscope frame (page 20). However, with the WI-DICD condenser, the DIC prism should be attached after adjusting the polarizer position.

1. Rotate the condenser height adjustment knob ② to raise the condenser holder to an optimum height.
2. Fully loosen the condenser clamping knob ③.
3. Slide in the condenser ④ from the front along the mount dovetail all the way until it is stopped.

★**When the microscope frame has a positioning pin on the rear position of the condenser, align the condenser with the groove on the condenser holder.**

4. Tighten the condenser clamping knob and lower the condenser holder to the lowest limit position.

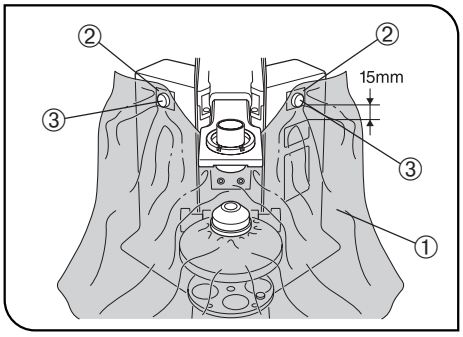


Fig. 50

5 Attaching the Waterproof Cover (Fig. 50)

◎Attach the waterproof cover onto the condenser if required.
The waterproof cover is applicable only to the WI-UCD, WI-DICD and WI-OBCD condensers.

1. Fit the hole of the waterproof cover ① in the extremity of the condenser and clamp with the clamping band.

CAUTION **In DIC observation, the condenser has to be removed and attached during adjustments. Therefore, in this case, do not attach the clamping band but just fit the hole of the waterproof cover in the extremity of the condenser and attach the clamping band after completing the adjustments.**

2. To fix the skirt of the waterproof cover, attach the double-side adhesive tape of the magnetic support plates ② to both sides of the microscope frame.

◎The magnetic support plates ② are most effective when they are attached symmetrically at positions by about 15 mm above the BX51WI indications.

3. Fix the waterproof cover using magnets ③.

◎When the cross stage is used, the stage mounting screw holes (x 4 on the front and rear) are hidden by the waterproof cover. However, this does not pose problem because the screws can later be attached by passing through the waterproof cover.

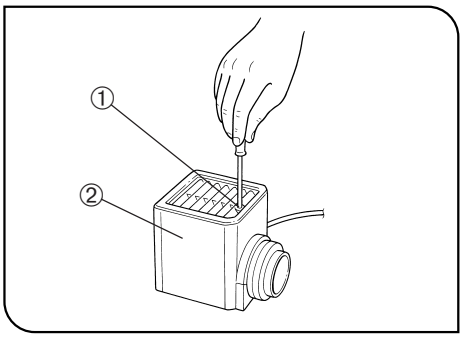


Fig. 51

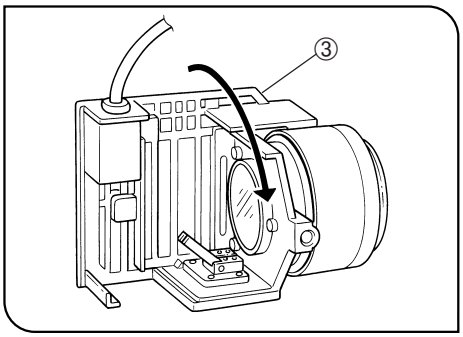


Fig. 52

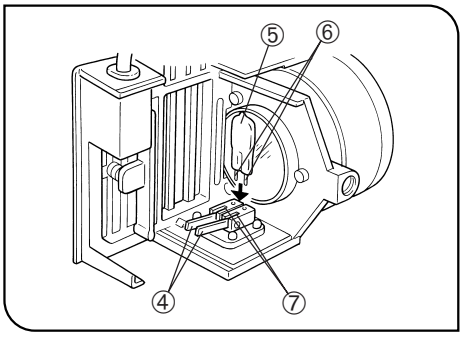


Fig. 53

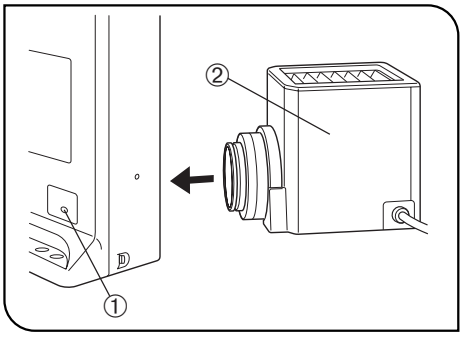


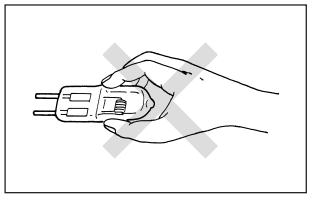
Fig. 54

6 Attaching the Halogen Lamp Housing (Figs. 51-54)

Attaching the Halogen Bulb

◎The applicable lamp bulb model is the 12V100WHAL-L (PHILIPS 7724) or the 12V50WHAL-L (LIFE JC).

1. Fully loosen the clamping screw ① at the top of the lamp housing using the Allen screwdriver provided with the microscope frame.
2. Remove the lamp housing ② by lifting it up.
3. Tilt the bulb socket ③ by 90° in the direction of the arrow.
4. While pushing down the bulb clamping lever ④ down, hold the halogen bulb ⑤ with gloves or a piece of gauze, insert the bulb pins ⑥ straight and fully into the sections ⑦ on the lamp socket.
   Then return the lamp clamping lever gently back to the original position to clamp the bulb.



▲To prevent reduced bulb life or cracking, do not touch the bulb with bare hands. If fingerprints are accidentally left on the bulb, wipe the bulb with a soft cloth.

5. Fit the lamp housing from up and tighten the clamping screw ① by applying downward pressure. (Fig. 51)

▲Caution for Bulb Replacement During or Right after Use

The bulb, lamp housing and areas near these will be extremely hot during and right after use.
Set the main switch to "O" (OFF), disconnect the power cord from the wall outlet, then allow the old bulb and lamp socket to cool before replacing the bulb with a new of the designated type.

Attaching the Halogen Lamp Housing

1. Loosen the bulb socket clamping screw ① using the Allen screwdriver.
2. Push in the halogen lamp housing ② with bulb and tighten the clamping screw ①.

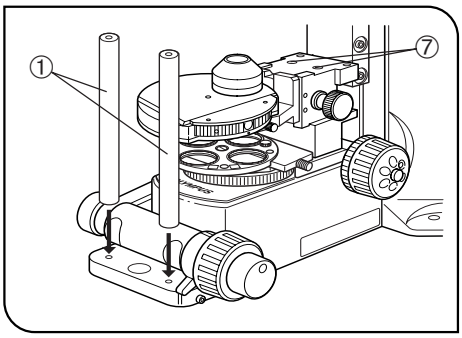


Fig. 55

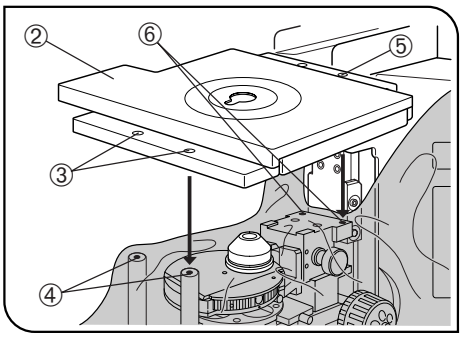


Fig. 56

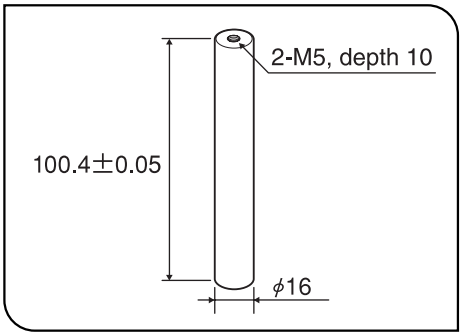


Fig. 57

7 Attaching the Cross Stage (Figs. 55 & 56)

◎When using a commercially marketed bridge stage, attach it by referring to its instruction manual.

1. Align the two WI-FSH fixed stage adapters ① with the front of the microscope base and clamp the adapters by tightening the hex-socket screws from the bottom side using the Allen wrench provided with the microscope frame.
2. Lower the condenser, align the mounting holes ③ and ⑤ of the IX-SVL2 cross stage ② with the mounting screw holes ④ and ⑥, and clamp the cross stage by tightening the hex-socket screws with the Allen wrench provided with the microscope frame.

◎When the waterproof cover is used, attach hex-socket screws to the screw holes ④ and ⑥ by passing through the waterproof cover.

Lowering the Stage Height

When no condenser is used, the stage height can be lowered by 50 mm by loosening the 2 condenser holder clamping screws (⑦ in Fig. 55) and removing the holder.

In this case, however, the length of the WI-FSH fixed stage adapter becomes excessive. To deal with this, order custom fabrication of two support pillars as shown in Fig. 57 or fabricate them by yourself.

PROPER SELECTION OF THE POWER SUPPLY CORD

If no power supply cord is provided, please select the proper power supply cord for the equipment by referring to " Specifications " and " Certified Cord " below:

CAUTION: In case you use a non-approved power supply cord for Olympus products, Olympus can no longer warrant the electrical safety of the equipment.

Specifications

Voltage Rating	125V AC (for 100-120V AC area) or, 250V AC (for 220-240V AC area)
Current Rating	6A minimum
Temperature Rating	60°C minimum
Length	3.05 m maximum
Fittings Configuration	Grounding type attachment plug cap. Opposite terminates in molded-on IEC configuration appliance coupling.

Table 1 Certified Cord

A power supply cord should be certified by one of the agencies listed in Table 1 , or comprised of cordage marked with an agency marking per Table 1 or marked per Table 2. The fittings are to be marked with at least one of agencies listed in Table 1. In case you are unable to buy locally in your country the power supply cord which is approved by one of the agencies mentioned in Table 1, please use replacements approved by any other equivalent and authorized agencies in your country.

Country	Agency	Certification Mark	Country	Agency	Certification Mark
Argentina	IRAM		Italy	IMQ	
Australia	SAA		Japan	JET, JQA, TÜV, UL-APEX / MITI	,
Austria	ÖVE		Netherlands	KEMA	
Belgium	CEBEC		Norway	NEMKO	
Canada	CSA		Spain	AEE	
Denmark	DEMKO		Sweden	SEMKO	
Finland	FEI		Switzerland	SEV	
France	UTE		United Kingdom	ASTA BSI	,
Germany	VDE		U.S.A.	UL	
Ireland	NSAI				

Table 2 HAR Flexible Cord

APPROVAL ORGANIZATIONS AND CORDAGE HARMONIZATION MARKING METHODS

Approval Organization	Printed or Embossed Harmonization Marking (May be located on jacket or insulation of internal wiring)		Alternative Marking Utilizing Black-Red-Yellow Thread (Length of color section in mm)		
			Black	Red	Yellow
Comite Electrotechnique Belge (CEBEC)	CEBEC	‹HAR›	10	30	10
Verband Deutscher Elektrotechniker (VDE) e.V. Prüfstelle	‹VDE›	‹HAR›	30	10	10
Union Technique de l´Electricite´ (UTE)	USE	‹HAR›	30	10	30
Instituto Italiano del Marchio di Qualita´ (IMQ)	IEMMEQU	‹HAR›	10	30	50
British Approvals Service for Electric Cables (BASEC)	BASEC	‹HAR›	10	10	30
N.V. KEMA	KEMA-KEUR	‹HAR›	10	30	30
SEMKO AB Svenska Elektriska Materielkontrollanstalter	SEMKO	‹HAR›	10	10	50
Österreichischer Verband für Elektrotechnik (ÖVE)	‹ÖVE›	‹HAR›	30	10	50
Danmarks Elektriske Materialkontroll (DEMKO)	‹DEMKO›	‹HAR›	30	10	30
National Standards Authority of Ireland (NSAI)	‹NSAI›	‹HAR›	30	30	50
Norges Elektriske Materiellkontroll (NEMKO)	NEMKO	‹HAR›	10	10	70
Asociacion Electrotecnica Y Electronica Espanola (AEE)	‹UNED›	‹HAR›	30	10	70
Hellenic Organization for Standardization (ELOT)	ELOT	‹HAR›	30	30	70
Instituto Portages da Qualidade (IPQ)	np	‹HAR›	10	10	90
Schweizerischer Elektro Technischer Verein (SEV)	SEV	‹HAR›	10	30	90
Elektriska Inspektoratet	SETI	‹HAR›	10	30	90

Underwriters Laboratories Inc. (UL)	SV, SVT, SJ or SJT, 3 X 18AWG
Canadian Standards Association (CSA)	SV, SVT, SJ or SJT, 3 X 18AWG

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